Appendices to the Biological and Water Quality Study of the Minor Great Black Swamp Tributaries, 2015-2016

Defiance, Fulton, Hancock, Henry, Paulding, Putnam, and Wood Counties, Ohio

Division of Surface Water
Ecological Assessment Section
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Appendices to the
Defiance, Fulton, Hancock, Henry, Paulding, Putnam, and Wood Counties, Ohio


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Notice to Users
NOTICE TO USERS

Ohio EPA incorporated biological criteria into the Ohio Water Quality Standards (WQS; Ohio Administrative Code 3745-1) regulations in February 1990 (effective May 1990). These criteria consist of numeric values for the Index of Biotic Integrity (IBI) and Modified Index of Well-Being (MIwb), both of which are based on fish assemblage data, and the Invertebrate Community Index (ICI), which is based on macroinvertebrate assemblage data. Criteria for each index are specified for each of Ohio's five ecoregions (as described by Omernik and Gallant 1988), and are further organized by organism group, index, site type, and aquatic life use designation. These criteria, along with the existing chemical and whole effluent toxicity evaluation methods and criteria, figure prominently in the monitoring and assessment of Ohio’s surface water resources.

The following documents support the use of biological criteria by outlining the rationale for using biological information, the methods by which the biocriteria were derived and calculated, the field methods by which sampling must be conducted, and the process for evaluating results:


Since the publication of the preceding guidance documents, the following new publications by the Ohio EPA have become available. These publications should also be consulted as they represent the latest information and analyses used by the Ohio EPA to implement the biological criteria.


These documents and this report may be obtained by contacting:

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**FOREWORD**

*What is a Biological and Water Quality Survey?*

A biological and water quality survey, or “biosurvey”, is an interdisciplinary monitoring effort coordinated on a waterbody specific or watershed scale. This effort may involve a relatively simple setting focusing on one or two small streams, one or two principal stressors, and a handful of sampling sites or a much more complex effort including entire drainage basins, multiple and overlapping stressors, and tens of sites. Each year the Ohio EPA conducts biosurveys in 4-5 watersheds study areas with an aggregate total of 250-300 sampling sites.

The Ohio EPA employs biological, chemical, and physical monitoring and assessment techniques in biosurveys in order to meet three major objectives: 1) determine the extent to which use designations assigned in the Ohio Water Quality Standards (WQS) are either attained or not attained; 2) determine if use designations assigned to a given water body are appropriate and attainable; and 3) determine if any changes in key ambient biological, chemical, or physical indicators have taken place over time, particularly before and after the implementation of point source pollution controls or best management practices. The data gathered by a biosurvey is processed, evaluated, and synthesized in a biological and water quality report. Each biological and water quality study contains a summary of major findings and recommendations for revisions to WQS, future monitoring needs, or other actions which may be needed to resolve existing impairment of designated uses. While the principal focus of a biosurvey is on the status of aquatic life uses, the status of other uses such as recreation and water supply, as well as human health concerns are also addressed.

The findings and conclusions of a biological and water quality study may factor into regulatory actions taken by the Ohio EPA (e.g., NPDES permits, Director’s Orders, the Ohio Water Quality Standards [OAC 3745-1], Water Quality Permit Support Documents [WQPSDs]), and are eventually incorporated into State Water Quality Management Plans, the Ohio Nonpoint Source Assessment, and the biennial Integrated Water Quality Monitoring and Assessment Report (305[b] and 303[d]).

*Hierarchy of Indicators*

A carefully conceived ambient monitoring approach, using cost-effective indicators consisting of ecological, chemical, and toxicological measures, can ensure that all relevant pollution sources are judged objectively on the basis of environmental results. Ohio EPA relies on a tiered approach in attempting to link the results of administrative activities with true environmental measures. This integrated approach includes a hierarchical continuum from administrative to true environmental indicators (Figure 1). The six “levels” of indicators include: 1) actions taken by regulatory agencies (permitting, enforcement, grants); 2) responses by the regulated community (treatment works, pollution prevention); 3) changes in discharged quantities (pollutant loadings); 4) changes in ambient conditions (water quality, habitat); 5) changes in uptake and/or assimilation (tissue contamination, biomarkers, wasteload allocation); and, 6) changes in health, ecology, or other effects (ecological condition, pathogens). The results of administrative activities (levels 1 and 2) can be linked to efforts to improve water quality (levels 3, 4, and 5) which should translate into the environmental “results” (level 6). Thus, the aggregate effect of billions of dollars spent on water pollution control since the early 1970s can now be determined with quantifiable measures of environmental condition.

Superimposed on this hierarchy is the concept of stressor, exposure, and response indicators. *Stressor* indicators generally include activities which have the potential to degrade the aquatic environment such as pollutant discharges (permitted and unpermitted), land use effects, and habitat modifications. *Exposure* indicators are those which measure the effects of stressors and can include whole effluent toxicity tests, tissue residues, and biomarkers, each of which provides evidence of biological exposure to
a stressor or bioaccumulative agent. *Response* indicators are generally composite measures of the cumulative effects of stress and exposure and include the more direct measures of community and population response that are represented here by the biological indices which comprise Ohio’s biological criteria. Other response indicators could include target assemblages, *i.e.*, rare, threatened, endangered, special status, and declining species or bacterial levels which serve as surrogates for the recreational uses. These indicators represent the essential technical elements for watershed-based management approaches. The key, however, is to use the different indicators *within* the roles which are most appropriate for each.

Describing the causes and sources associated with observed impairments revealed by the biological criteria and linking this with pollution sources involves an interpretation of multiple lines of evidence including water chemistry data, sediment data, habitat data, effluent data, biomonitoring results, land use data, and biological response signatures within the biological data itself. Thus the assignment of principal causes and sources of impairment represents the association of impairments (defined by response indicators) with stressor and exposure indicators. The principal reporting venue for this process on a watershed or subbasin scale is a biological and water quality report. These reports then provide the foundation for aggregated assessments such as the Integrated Report, the Ohio Nonpoint Source Assessment, and other technical bulletins.
Ohio Water Quality Standards: Designated Aquatic Life Use

The Ohio Water Quality Standards (WQS; Ohio Administrative Code 3745-1) consist of designated uses and chemical, physical, and biological criteria designed to represent measurable properties of the environment that are consistent with the goals specified by each use designation. Use designations consist of two broad groups, aquatic life and non-aquatic life uses. In applications of the Ohio WQS to the management of water resource issues in Ohio’s rivers and streams, the aquatic life use criteria frequently result in the most stringent protection and restoration requirements, hence their emphasis in biological and water quality reports. Also, an emphasis on protecting for aquatic life generally results in water quality suitable for all uses. The five different aquatic life uses currently defined in the Ohio WQS are described as follows:

1) **Warmwater Habitat (WWH)** - this use designation defines the “typical” warmwater assemblage of aquatic organisms for Ohio rivers and streams; *this use represents the principal restoration target for the majority of water resource management efforts in Ohio.*

2) **Exceptional Warmwater Habitat (EWH)** - this use designation is reserved for waters which support “unusual and exceptional” assemblages of aquatic organisms which are characterized by a high diversity of species, particularly those which are highly intolerant and/or rare, threatened, endangered, or special status (i.e., declining species); *this designation represents a protection goal for water resource management efforts dealing with Ohio’s best water resources.*

3) **Coldwater Habitat (CWH)** - this use is intended for waters which support assemblages of coldwater organisms and/or those which are stocked with salmonids with the intent of providing a put-and-take fishery on a year round basis which is further sanctioned by the Ohio DNR, Division of Wildlife; this use should not be confused with the Seasonal Salmonid Habitat (SSH) use which applies to the Lake Erie tributaries which support periodic “runs” of salmonids during the spring, summer, and/or fall.

4) **Modified Warmwater Habitat (MWH)** - this use applies to streams and rivers which have been subjected to extensive, maintained, and essentially permanent hydromodifications such that the biocriteria for the WWH use are not attainable and *where the activities have been sanctioned by state or federal law*; the representative aquatic assemblages are generally composed of species which are tolerant to low dissolved oxygen, silt, nutrient enrichment, and poor quality habitat.

5) **Limited Resource Water (LRW)** - this use applies to small streams (usually <3 mi² drainage area) and other water courses which have been irretrievably altered to the extent that no appreciable assemblage of aquatic life can be supported; such waterways generally include small streams in extensively urbanized areas, those which lie in watersheds with extensive drainage modifications, those which completely lack water on a recurring annual basis (i.e., true ephemeral streams), or other irretrievably altered waterways.

Chemical, physical, and/or biological criteria are generally assigned to each use designation in accordance with the broad goals defined by each. As such the system of use designations employed in the Ohio WQS constitutes a “tiered” approach in that varying and graduated levels of protection are provided by each. This hierarchy is especially apparent for parameters such as dissolved oxygen, ammonia-nitrogen, temperature, and the biological criteria. For other parameters such as heavy metals,
the technology to construct an equally graduated set of criteria has been lacking, thus the same WQS criteria may apply to two or three different use designations.

Ohio Water Quality Standards: Non-Aquatic Life Uses
In addition to assessing the appropriateness and status of aquatic life uses, each biological and water quality survey also addresses non-aquatic life uses such as recreation, water supply, and human health concerns as appropriate. The recreation uses most applicable to rivers and streams are the Primary Contact Recreation (PCR) and Secondary Contact Recreation (SCR) uses. The criterion for designating the PCR use can be having a water depth of at least one meter over an area of at least 100 square feet or, lacking this, where frequent human contact is a reasonable expectation. If a water body does not meet either criterion, the SCR use applies. The attainment status of PCR and SCR is determined using bacterial indicators (e.g., fecal coliform, E. coli) and the criteria for each are specified in the Ohio WQS.

Attainment of recreation uses are evaluated based on monitored bacteria levels. The Ohio Water Quality Standards state that all waters should be free from any public health nuisance associated with raw or poorly treated sewage (Administrative Code 3745-1-04, Part F). Additional criteria (Administrative Code 3745-1-07) apply to waters that are designated as suitable for full body contact such as swimming (PCR) or for partial body contact such as wading (SCR). These standards were developed to protect human health, because even though fecal coliform bacteria are relatively harmless in most cases, their presence indicates that the water has been contaminated with fecal matter.

Water supply uses include Public Water Supply (PWS), Agricultural Water Supply (AWS), and Industrial Water Supply (IWS). Public Water Supplies are simply defined as segments within 500 yards of a potable water supply or food processing industry intake. The Agricultural Water Supply (AWS) and Industrial Water Supply (IWS) use designations generally apply to all waters unless it can be clearly shown that they are not applicable. An example of this would be an urban area where livestock watering or pasturing does not take place, thus the AWS use would not apply. Chemical criteria are specified in the Ohio WQS for each use and attainment status is based primarily on chemical-specific indicators. Human health concerns are additionally addressed with fish tissue data, but any consumption advisories are issued by the Ohio Department of Health.
MECHANISMS FOR WATER QUALITY IMPAIRMENT

The following paragraphs are provided to present the varied causes of impairment that affect the resource quality of lotic systems in Ohio. While the various perturbations are presented under separate headings, it is important to remember that they are often interrelated and cumulative in terms of the detrimental impact that can result.

Habitat and Flow Alterations
Habitat alteration, such as channelization, negatively impacts biological communities directly by limiting the complexity of living spaces available to aquatic organisms. Consequently, fish and macroinvertebrate communities are not as diverse. Indirect impacts include the removal of riparian trees and field tiling to facilitate drainage. Following a rain event, most of the water is quickly removed from tiled fields rather than filtering through the soil, recharging ground water, and reaching the stream at a lower volume and more sustained rate. As a result, small streams more frequently go dry or become intermittent. Urbanization impacts include removal of riparian trees, influx of storm water runoff, straightening and piping of stream channels, and riparian vegetation removal.

Tree shade is important because it limits the energy input from the sun, moderates water temperature, and limits evaporation. Removal of the tree canopy further degrades conditions because it eliminates an important source of coarse organic matter essential for a balanced ecosystem. Riparian vegetation aids in nutrient uptake, may decrease runoff rate into streams, and helps keep soil in place. Erosion impacts channelized streams more severely due to the lack of a riparian buffer zone to slow runoff, trap sediment and stabilize banks. Additionally, deep trapezoidal channels lack a functioning flood plain and therefore cannot expel sediment as would occur during flood events along natural watercourses. The confinement of flow within an artificially deep channel accelerates the movement of water downstream, exacerbating flooding of neighboring properties.

The lack of water movement under low flow conditions can exacerbate impacts from organic loading and nutrient enrichment by limiting re-aeration of the stream. The amount of oxygen soluble in water decreases as temperature increases. This is one reason why tree shade is so important. The two main sources of oxygen in water are diffusion from the atmosphere and plant photosynthesis. Turbulence at the water surface is critical because it increases surface area and promotes diffusion, but channelization eliminates turbulence produced by riffles, meanders, and debris snags. Plant photosynthesis produces oxygen, but at night, respiration reverses the process and consumes oxygen. Conversely, oxygen concentrations can become supersaturated during the day, due to abnormally high amounts of photosynthesis, causing gas bubble stress to both fish and invertebrate communities. Oxygen is also used by bacteria that decay dead organic matter. Nutrient enrichment can promote the growth of nuisance algae that subsequently dies and serves as food for bacteria. Under these conditions, oxygen can be depleted unless it is replenished from the air.

Siltation and Sedimentation
Whenever the natural flow regime is altered to facilitate drainage, increased amounts of sediment are likely to enter streams either by overland transport or increased bank erosion. The removal of wooded riparian areas furthers the erosion process. Channelization keeps all but the highest flow events confined within the artificially high banks. As a result, areas that were formerly flood plains and allowed for the removal of sediment from the primary stream channel no longer serve this function. As water
levels fall following a rain event, interstitial spaces between larger rocks fill with sand and silt and the diversity of available habitat to support fish and macroinvertebrates is reduced. Silt also can clog the gills of both fish and macroinvertebrates, reduce visibility thereby excluding site feeding fish species, and smother the nests of lithophilic fishes. Lithophilic spawning fish require clean substrates with interstitial voids in which to deposit eggs. Conversely, pioneering species benefit. They are generalists and best suited for exploiting disturbed and less heterogeneous habitats. The net result is a lower diversity of aquatic species compared with a typical warmwater stream with natural habitats.

Sediment also impacts water quality, recreation, and drinking water. Nutrients absorbed to soil particles remain trapped in the watercourse. Likewise, bacteria, pathogens, and pesticides which also attach to suspended or bedload sediments become concentrated in waterways where the channel is functionally isolated from the landscape. Community drinking water systems address these issues with more costly advanced treatment technologies.

**Nutrient Enrichment**

The element of greatest concern is phosphorus because it is critical for plant growth and is often the limiting nutrient. The form that can be readily used by plants and therefore can stimulate nuisance algae blooms is orthophosphate (PO$_4^{3-}$). The amount of phosphorus tied up in the nucleic acids of food and waste is actually quite low. This organic material is eventually converted to orthophosphate by bacteria. The amount of orthophosphate contained in synthetic detergents is a great concern however. It was for this reason that the General Assembly of the State of Ohio enacted a law in 1990 to limit phosphorus content in household laundry detergents sold in the Lake Erie drainage basin to 0.5% by weight. Inputs of phosphorus originate from both point and nonpoint sources. Most of the phosphorus discharged by point sources is soluble. Another characteristic of point sources is they have a continuous impact and are human in origin, for instance, effluents from municipal sewage treatment plants. The contribution from failed on-lot septic systems can also be significant, especially if they are concentrated in a small area. The phosphorus concentration in raw waste water is generally 8-10 mg/l and after secondary treatment is generally 4-6 mg/l. Further removal requires the added cost of chemical addition. The most common methods use the addition of lime or alum to form a precipitate, so most phosphorus (80%) ends up in the sludge.

A characteristic of phosphorus discharged by nonpoint sources is that the impact is intermittent and associated with storm water runoff. Most of this phosphorus is bound tightly to soil particles and enters streams from erosion, although some comes from tile drainage. Urban storm water is more of a concern if combined sewer overflows are involved. The impact from rural storm water varies depending on land use and management practices and includes contributions from livestock feedlots and pastures and row crop agriculture. Crop fertilizer includes granular inorganic types and organic types such as manure or sewage sludge. Pasture land is especially a concern if the livestock have access to the stream. Large feedlots with manure storage lagoons create the potential for overflows and accidental spills. Land management is an issue because erosion is worse on streams without any riparian buffer zone to trap runoff. The impact is worse in streams that are channelized because they no longer have a functioning flood plain and cannot expel sediment during flooding. Oxygen levels must also be considered, because phosphorus is released from sediment at higher rates under anoxic conditions.

There is no numerical phosphorus criterion established in the Ohio Water Quality Standards, but there is a narrative criterion that states phosphorus should be limited to the extent necessary to prevent nuisance growths of algae and weeds (Administrative Code, 3745-1-04, Part E). Phosphorus loadings from large volume point source dischargers in the Lake Erie drainage basin are regulated by NPDES permit limits. The permit limit is a concentration of 1.0 mg/l in final effluent. Research conducted by the Ohio EPA indicates that a significant correlation exists between phosphorus and the health of aquatic communities (Miltner and Rankin, 1998). It was concluded that biological community performance in
headwater and wadeable streams was highest where phosphorus concentrations were lowest. It was also determined that the lowest phosphorus concentrations were associated with the highest quality habitats, supporting the notion that habitat is a critical component of stream function. The report recommends WWH total phosphorus targets of 0.08 mg/l in headwater streams (<20 mi² watershed size), 0.10 mg/l in wadeable streams (>20-200 mi²) and 0.17 mg/l in small rivers (>200-1000 mi²).

Organic Enrichment and Low Dissolved Oxygen
The amount of oxygen soluble in water is low and it decreases as temperature increases. This is one reason why tree shade is so important. The two main sources of oxygen in water are diffusion from the atmosphere and plant photosynthesis. Turbulence at the water surface is critical because it increases surface area and promotes diffusion. Drainage practices such as channelization eliminate turbulence produced by riffles, meanders, and debris snags. Although plant photosynthesis produces oxygen by day, it is consumed by the reverse process of respiration at night. Oxygen is also consumed by bacteria that decay organic matter, so it can be easily depleted unless it is replenished from the air. Sources of organic matter include poorly treated waste water, sewage bypasses, and dead plants and algae. Dissolved oxygen criteria are established in the Ohio Water Quality Standards to protect aquatic life. The minimum and average limits are tiered values and linked to use designations (Administrative Code 3745-1-07, Table 7-1).

Ammonia
Ammonia enters streams as a component of fertilizer and manure run-off and wastewater effluent. Ammonia gas (NH₃) readily dissolves in water to form the compound ammonium hydroxide (NH₃OH). In aquatic ecosystems, equilibrium is established as ammonia shifts from a gas to undissociated ammonium hydroxide to the dissociated ammonium ion (NH₄+). Under normal conditions (neutral pH 7.0 and temperature 25°C), almost none of the total ammonia is present as gas, only 0.55% is present as ammonium hydroxide, and the rest is ammonium ion. Alkaline pH shifts the equation toward gaseous ammonia production, so the amount of ammonium hydroxide increases. This is important because while the ammonium ion is almost harmless to aquatic life, ammonium hydroxide is very toxic and can reduce growth and reproduction or cause mortality.

The concentration of ammonia in raw sewage is high, sometimes as much as 20-30 mg/l. Treatment to remove ammonia involves gaseous stripping to the atmosphere, biological nitrification and denitrification, and assimilation into plant and animal biomass. The nitrification process requires a long detention time and aerobic conditions like that provided in extended aeration treatment plants. Under these conditions, bacteria first convert ammonia to nitrite (Nitrosomonas) and then to nitrate (Nitrobacter). Nitrate can then be reduced by the de-nitrification process (Pseudomonas) and nitrogen gas and carbon dioxide are produced as by-products.

Ammonia criteria are established in the Ohio Water Quality Standards to protect aquatic life. The maximum and average limits are tiered values based on sample pH and temperature and linked to use designations (Administrative Code 3745-1-07, Tables 7-2 through 7-8).

Metals
Metals can be toxic to aquatic life and hazardous to human health. Although they are naturally occurring elements many are extensively used in manufacturing and are byproducts of human activity. Certain metals like copper and zinc are essential in the human diet, but excessive levels are usually detrimental. Lead and mercury are of particular concern because they often trigger fish consumption advisories. Mercury is used in the production of chlorine gas and caustic soda and in the manufacture of batteries and fluorescent light bulbs. In the environment it forms inorganic salts, but bacteria convert these to methyl-mercury and this organic form builds up in the tissues of fish. Extended exposure can damage
the brain, kidneys, and developing fetus. The Ohio Department of Health (ODH) issued a statewide fish consumption advisory in 1997 advising women of child bearing age and children six and under not to eat more than one meal per week of any species of fish from waters of the state because of mercury. Lead is used in batteries, pipes, and paints and is emitted from burning fossil fuels. It affects the central nervous system and damages the kidneys and reproductive system. Copper is mined extensively and used to manufacture wire, sheet metal, and pipes. Ingesting large amounts can cause liver and kidney damage. Zinc is a by-product of mining, steel production, and coal burning and used in alloys such as brass and bronze. Ingesting large amounts can cause stomach cramps, nausea, and vomiting.

Metals criteria are established in the Ohio Water Quality Standards to protect human health, wildlife, and aquatic life. Three levels of aquatic life standards are established (Administrative Code 3745-1-07, Table 7-1) and limits for some elements are based on water hardness (Administrative Code 3745-1-07, Table 7-9). Human health and wildlife standards are linked to either the Lake Erie (Administrative Code 3745-1-33, Table 33- 2) or Ohio River (Administrative Code 3745-1-34, Table 34-1) drainage basins. The drainage basins also have limits for additional elements not established elsewhere that are identified as Tier I and Tier II values.

**Bacteria**

High concentrations of either fecal coliform bacteria or *Escherichia coli* (*E. coli*) in a lake or stream may indicate contamination with human pathogens. People can be exposed to contaminated water while wading, swimming, and fishing. Fecal coliform bacteria are relatively harmless in most cases, but their presence indicates that the water has been contaminated with feces from a warm-blooded animal. Although intestinal organisms eventually die off outside the body, some will remain virulent for a period of time and may be dangerous sources of infection. This is especially a problem if the feces contained pathogens or disease producing bacteria and viruses. Reactions to exposure can range from an isolated illness such as skin rash, sore throat, or ear infection to a more serious wide spread epidemic. Some types of bacteria that are a concern include *Escherichia*, which cause diarrhea and urinary tract infections, *Salmonella*, which cause typhoid fever and gastroenteritis (food poisoning), and *Shigella*, which cause severe gastroenteritis or bacterial dysentery. Some types of viruses that are a concern include polio, hepatitis A, and encephalitis. Disease causing microorganisms such as cryptosporidium and giardia are also a concern.

Since fecal coliform bacteria are associated with warm-blooded animals, there are both human and animal sources. Human sources, including effluent from sewage treatment plants or discharges by on-lot septic systems, are a more continuous problem. Bacterial contamination from combined sewer overflows are associated with wet weather events. Animal sources are usually more intermittent and are also associated with rainfall, except when domestic livestock have access to the water. Large livestock farms store manure in holding lagoons and this creates the potential for an accidental spill. Liquid manure applied as fertilizer is a runoff problem if not managed properly and it sometimes seeps into field tiles.

Bacteria criteria for the recreational use are established in the Ohio Water Quality Standards to protect human health. The maximum and average limits are tiered *E. coli* values and linked to use designation, but only apply during the May 1-October 15 recreation season (Administrative Code 3745-1-07, Table 7-13). The standards also state that streams must be free of any public health nuisance associated with raw or poorly treated sewage during dry weather conditions (Administrative Code 3745-1-04, Part F).
Sediment Contamination

Chemical quality of sediment is a concern because many pollutants bind strongly to soil particles and are persistent in the environment. Some of these compounds accumulate in the aquatic food chain and trigger fish consumption advisories, but others are simply a contact hazard because they cause skin cancer and tumors. The physical and chemical nature of sediment is determined by local geology, land use, and contribution from manmade sources. As some materials enter the water column they are attracted to the surface electrical charges associated with suspended silt and clay particles. Others simply sink to the bottom due to their high specific gravity. Sediment layers form as suspended particles settle, accumulate, and combine with other organic and inorganic materials. Sediment is the most physically, chemically, and biologically reactive at the water interface because this is where it is affected by sunlight, current, wave action, and benthic organisms. Assessment of the chemical nature of this layer can be used to predict ecological impact.

Sediment data are evaluated using Ohio Sediment Reference Values (SRVs; Ohio EPA 2008), along with guidelines established in Development and Evaluation of Consensus-Based Sediment Quality Guidelines for Freshwater Ecosystems (MacDonald et al. 2000), and Ecological Screening Levels (ESLs) (U.S. EPA 2003). The Ohio EPA system was derived from samples collected at ecoregional reference sites. Specific SRVs are site specific ecoregional based metals concentrations and are used to identify contaminated stream reaches. The MacDonald guidelines are consensus based using previously developed values. The system predicts that sediments below the threshold effect concentration (TEC) are absent of toxicity and those greater than the probable effect concentration (PEC) are toxic. ESL values, considered protective benchmarks, were derived by US EPA Region 5 using a variety of sources and methods.

Sediment samples collected by the Ohio EPA are measured for a number of physical and chemical properties. Physical attributes included percent particle size distribution (sand ≥60μ, silt 5-59μ, clay ≤4μ), percent solids, and percent organic carbon. Most locations sampled had an abundance of sediment, and no difficulties were experienced in locating ample volumes of sediment for analysis. Fine grained sediments are deposited in flood plains of natural streams during periods of high flow. This scenario changes if the stream is impounded by a dam or channelized. Chemical attributes included metals, volatile and semi-volatile organic compounds, pesticides, and polychlorinated biphenyls (PCBs).

MATERIALS and METHODS

All biological, chemical, and physical habitat data collection, processing, and analysis methods and procedures adhere to those specified in the Surface Water Field Sampling Manual for water column chemistry, bacteria and flows (Ohio EPA 2013), Biological Criteria for the Protection of Aquatic Life, Volumes II - III (Ohio EPA 1987b, 1989a, 1989b, 2015a, 2015b), and The Qualitative Habitat Evaluation Index (QHEI): Rationale, Methods, and Application (Rankin 1989).

Determining Use Attainment Status

Use attainment status is a term describing the degree to which environmental indicators are either above or below criteria specified by the Ohio Water Quality Standards (WQS; Ohio Administrative Code 3745-1). Assessing aquatic use attainment status involves a primary reliance on the Ohio EPA biological criteria (OAC 3745-1-07; Table 7-15). These are confined to ambient assessments and apply to rivers and streams outside of mixing zones. Numerical biological criteria are based on multimetric biological indices including the IBI and MIwb, indices measuring the response of the fish community, and the ICI, which indicates the response of the macroinvertebrate community. Three attainment status results are possible at each sampling location - full, partial, or non-attainment. Full attainment means that all of the applicable indices meet the biocriteria. Partial attainment means that one or more of the applicable indices fails to meet the biocriteria. Non-attainment means that none of the applicable indices meet the
biocriteria or one of the organism groups reflects poor or very poor performance. An aquatic life use attainment table is constructed based on the sampling results and is arranged from upstream to downstream and includes the sampling locations indicated by river mile, the applicable biological indices, the use attainment status (i.e., full, partial, or non), the Qualitative Habitat Evaluation Index (QHEI), and a sampling location description.

**Habitat Assessment**
Physical habitat was evaluated using the QHEI developed by the Ohio EPA for streams and rivers in Ohio (Rankin 1989 and 1995, Ohio EPA 2006). Various attributes of the habitat are scored based on the overall importance of each to the maintenance of viable, diverse, and functional aquatic faunas. The type(s) and quality of substrates, amount and quality of instream cover, channel morphology, extent and quality of riparian vegetation, pool, run, and riffle development and quality, and gradient are some of the habitat characteristics used to determine the QHEI score which generally ranges from 20 to less than 100. The QHEI is used to evaluate the characteristics of a stream segment, as opposed to the characteristics of a single sampling site. As such, individual sites may have poorer physical habitat due to a localized disturbance yet still support aquatic communities closely resembling those sampled at adjacent sites with better habitat, provided water quality conditions are similar. QHEI scores from hundreds of segments around the state have indicated that values greater than 60 are generally conducive to the existence of warmwater faunas whereas scores less than 45 generally cannot support a warmwater assemblage consistent with the WWH biological criteria. Scores greater than 75 frequently reflect habitat conditions which have the ability to support exceptional warmwater faunas.

**Sediment and Surface Water Assessment**
Fine grain sediment samples were collected in the upper 4 inches of bottom material at each location using decontaminated stainless steel scoops and excavated using nitrile gloves. Decontamination of sediment sampling equipment followed the procedures outlined in the Ohio EPA sediment sampling guidance manual (Ohio EPA 2012). Sediment grab samples were homogenized in stainless steel pans (material for VOC analysis was not homogenized), transferred into glass jars with teflon® lined lids, placed on ice (to maintain 4°C) in a cooler, and shipped to Ohio EPA Division of Environmental Services. Sediment data is reported on a dry weight basis. Surface water samples were collected, preserved and delivered in appropriate containers to Ohio EPA Division of Environmental Services. Surface water samples were evaluated using comparisons to Ohio Water Quality Standards criteria, reference conditions, or published literature. Sediment evaluations were conducted using guidelines established in MacDonald et al. (2000), U.S. EPA (2003) and Ohio EPA (2008).

**Recreation Use Assessment**
Recreation use attainment was determined using the criteria established in OAC 3745-1-41:

1) *E. coli* is the only indicator organism used to evaluate recreation.
2) The recreation season extends from May 1 – Oct. 31.
3) Geometric mean content is computed on a seasonal basis.
4) Geometric mean content is the sole basis of use attainment status when 2 or more samples are taken.
5) Primary Contact Recreation (PCR) includes three separate categories each with specific numerical criteria: Class A – high use paddling streams, Class B – most typical streams and Class C – historically channelized streams that drain < 3.1 mi².

**Macroinvertebrate Community Assessment**
Macroinvertebrates were collected from artificial substrates and from the natural habitats. The artificial substrate collection provided quantitative data and consisted of a composite sample of five modified Hester-Dendy multiple-plate samplers colonized for six weeks. At the time of the artificial substrate
collection, a qualitative multihabitat composite sample was also collected. This sampling effort consisted of an inventory of all observed macroinvertebrate taxa from the natural habitats at each site with no attempt to quantify populations other than notations on the predominance of specific taxa or taxa groups within major macrohabitat types (e.g., riffle, run, pool, margin). Detailed discussion of macroinvertebrate field and laboratory procedures is contained in Biological Criteria for the Protection of Aquatic Life: Volume III, Standardized Biological Field Sampling and Laboratory Methods for Assessing Fish and Macroinvertebrate Communities (Ohio EPA 1989b, 2015b).

**Fish Community Assessment**

Fish were sampled using pulsed DC electrofishing methods. Fish were processed in the field, and included identifying each individual to species, counting, weighing, and recording any external abnormalities. Discussion of the fish community assessment methodology used in this report is contained in Biological Criteria for the Protection of Aquatic Life: Volume III, Standardized Biological Field Sampling and Laboratory Methods for Assessing Fish and Macroinvertebrate Communities (Ohio EPA 1989b, 2015b).

**Causal Associations**

Using the results, conclusions, and recommendations of this report requires an understanding of the methodology used to determine the use attainment status and assigning probable causes and sources of impairment. The identification of impairment in rivers and streams is straightforward - the numerical biological criteria are used to judge aquatic life use attainment and impairment (partial and non-attainment). The rationale for using the biological criteria, within a weight of evidence framework, has been extensively discussed elsewhere (Karr *et al.* 1986; Karr 1991; Ohio EPA 1987a, Ohio EPA 1987b; Yoder 1989; Miner and Borton 1991; Yoder 1991; Yoder 1995). Describing the causes and sources associated with observed impairments relies on an interpretation of multiple lines of evidence including water chemistry data, sediment data, habitat data, effluent data, land use data, and biological results (Yoder and Rankin 1995a, 1995b, and 1995c). Thus the assignment of principal causes and sources of impairment in this report represent the association of impairments (based on response indicators) with stressor and exposure indicators. The reliability of the identification of probable causes and sources is increased where many such prior associations have been identified, or have been experimentally or statistically linked together. The ultimate measure of success in water resource management is the restoration of lost or damaged ecosystem attributes including aquatic community structure and function. While there have been criticisms of misapplying the metaphor of ecosystem “health” compared to human patient “health” (Suter 1993), in this document we are referring to the process for evaluating biological integrity and causes or sources associated with observed impairments, not whether human health and ecosystem health are analogous concepts.
References


____1989a. Addendum to biological criteria for the protection of aquatic life: Volume II. Users manual for biological field assessment of Ohio surface waters. Division of Water Quality Planning and Assessment, Surface Water Section, Columbus, Ohio.

____1989b. Biological criteria for the protection of aquatic life: Volume III. Standardized biological field sampling and laboratory methods for assessing fish and macroinvertebrate communities. Division of Water Quality Planning and Assessment, Columbus, Ohio.


Appendix A

Macroinvertebrate Collection Results
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W. BR. TONTAGONY CREEK NEAR CEMETARY AT RANGELINE RD.
Rivercode: 04-013-001
River Mile: 2.19
0:00 8/29/16

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<tr>
<td>22300 Argia sp</td>
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</tr>
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<tr>
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</tr>
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<td>Pyganodon grandis</td>
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**Taxa**

<p>| 01320 | Hydra sp | 33 | 85800 | Tanytarsus sp | + |
| 01801 | Turbellaria | 40+ | 85821 | Tanytarsus glabrescens group sp 7 | 31+ |
| 03360 | Plumatella sp | + | 85840 | Tanytarsus sepp | + |
| 05900 | Lirceus sp | + | 87540 | Hemerodromia sp | |
| 08601 | Hydrachnidia | 10+ | 89501 | Ephydridae | 41 |
| 11020 | Acerpenna pygmaea | 51+ | 95100 | Physella sp | + |
| 11130 | Baetis intercalaris | 178+ | 98600 | Sphaerium sp | + |
| 11670 | Procloeon viridoculare | + | 99100 | Pyganodon grandis | + |
| 12200 | Isonychia sp | 4+ | 99240 | Lamigona complanata | + |
| 13400 | Stenacron sp | 96+ | 99380 | Quadrula pustulosa pustulosa | + |
| 13521 | Stenonema femoratum | + | 99400 | Quadrula quadrula | + |
| 13570 | Macccaffertium terminatum | 436+ | 99440 | Fusconaia flava | + |
| 16700 | Tricorythodes sp | 10+ | 99660 | Truncilla truncata | + |
| 17200 | Caenis sp | + | 99680 | Leptodea fragilis | + |
| 21200 | Calopteryx sp | 1+ | 99700 | Potamilius alatus | + |
| 22001 | Coenagroniidae | + | 99860 | Lampsilis radiata luteola | + |
| 22300 | Argia sp | 17+ | | + | |
| 23909 | Boyeria vinosa | + | | + | |
| 42700 | Belostoma sp | + | | + | |
| 50315 | Chimarra obscura | + | | + | |
| 52200 | Cheumatopsyche sp | 1,279+ | | + | |
| 52430 | Ceratopsyche morosa group | + | | + | |
| 52570 | Hydropsyche simulans | 433+ | | + | |
| 53800 | Hydroptila sp | 9+ | | + | |
| 59300 | Mystacides sp | + | | + | |
| 59407 | Nectopsyche candida | + | | + | |
| 68300 | Cyphon sp | 74 | | + | |
| 68601 | Ancyronyx variegata | 1+ | | + | |
| 68708 | Dubravigia vittata group | + | | + | |
| 69400 | Stenelmis sp | + | | + | |
| 71910 | Tipula abdominalis | 200 | | + | |
| 72700 | Anopheles sp | 8+ | | + | |
| 74650 | Atrochopogon sp | 96 | | + | |
| 77100 | Ablabesmyia sp | 16 | | + | |
| 77120 | Ablabesmyia mallochi | + | | + | |
| 77750 | Thienemanninimia sp | 31 | | + | |
| 78450 | Nilotanytarsus fimbratus | 78 | | + | |
| 78655 | Procladius (Holotanytarsus) sp | + | | + | |
| 80370 | Corynoneura lobata | 32 | | + | |
| 81825 | Rheocricotopus (Psilocricotopus) robacki | 31 | | + | |
| 82121 | Thiemenanniella lobapodema | 8 | | + | |
| 82130 | Thiemenanniella similis | 34 | | + | |
| 82141 | Thiemenanniella xena | 10+ | | + | |
| 82730 | Chironomus (C.) decorus group | + | | + | |
| 82824 | Cryptochironomus ponderosus | + | | + | |
| 82885 | Cryptotendipes pseudotener | + | | + | |
| 83040 | Dicrotendipes neomodestus | 47+ | | + | |
| 84300 | Phaenopsectra obediens group | + | | + | |
| 84450 | Polypedilum (Uresipedium) flavum | 408+ | | + | |
| 84460 | Polypedilum (P.) fallax group | 16 | | + | |
| 84470 | Polypedilum (P.) ilinoense | 204+ | | + | |
| 84750 | Stictochironomus sp | + | | + | |
| 85230 | Cladotanytarsus mancerus group | + | | + | |
| 85500 | Paratanytarsus sp | 16 | | + | |
| 85615 | Rheotanytarsus pellucidus | 63 | | + | |
| 85625 | Rheotanytarsus sp | 518+ | | + | |</p>
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Rivercode: 04-018-000
River Mile: 8.99

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**Taxa**

- Turbellaria +
- Oligochaeta +
- Helobdella stagnalis +
- Placobdella ornata +
- Lirceus sp +
- Orconectes (Gremicambarus) immunis +
- Acerpenna pygmaea +
- Stenacron sp +
- Caenis sp +
- Hetaerina sp +
- Coenagrionidae +
- Argia sp +
- Boyeria vinosa +
- Gomphus sp +
- Corixidae +
- Cheumatopsyche sp +
- Ceratopsyche morosa group +
- Hydropsyche depravata group +
- Hydroptila sp +
- Helicopsyche borealis +
- Nectopsyche diarina +
- Oecetis nocturna +
- Peltodytes sp +
- Berosus sp +
- Dubraphia vittata group +
- Macrancyclus gabratus +
- Simulium sp +
- Conchapelopia sp +
- Thienemanniomyia (Hayesomyia) senata +
- Pentaneura inconspicua +
- Cricotopus (C.) bicinctus +
- Cricotopus (C.) tremulus group +
- Cricotopus (Isocladius) sylvestris group +
- Thienemanniella lobapodema +
- Thienemanniella xena +
- Chironomus (C.) decorus group +
- Cryptochironomus sp +
- Cryptochironomus ponderosus +
- Dicrotendipes neomodestus +
- Paralauterborniia nigrohalteralis +
- Polypedilum (Uresipedilum) flavum +
- Polypedilum (P.) illinoense +
- Tribelos jucundum +
- Pseudochironomus sp +
- Cladotanytarsus muncas group +
- Paratanytarsus sp +
- Rheotanytarsus sp +
- Fossaria sp +
- Physella sp +
- Gyraulus (Torquis) parvus +
- Planor bella (Pierosoma) trivolvis +
- Ferrissia sp +
- Laevapex fuscus +
- Pisidium sp +
- Sphaerium sp +
- Strophitus undulatus undulatus +
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<tr>
<td>01801 Turbellaria                                                    +</td>
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<tr>
<td>05900 Lirceus sp                                                    +</td>
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<td>08250 Orconectes (Procericambarus) rusticus                         +</td>
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<td>08601 Hydrachnidia                                                  +</td>
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<td>11018 Acerpenna macdunnoughi                                        +</td>
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<td>11020 Acerpenna pygmaea                                              +</td>
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<td>11120 Baetis flavistriga                                            +</td>
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<td>11125 Labiobaetis frondalis                                         +</td>
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<tr>
<td>11130 Baetis intercalaris                                           +</td>
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<tr>
<td>13000 Leucrocuta sp                                                 +</td>
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<tr>
<td>13400 Stenacron sp                                                  +</td>
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<tr>
<td>17200 Caenis sp                                                     +</td>
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<tr>
<td>21200 Calopteryx sp                                                 +</td>
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<tr>
<td>23909 Boyeria vinosa                                                +</td>
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<tr>
<td>24107 Nasiaeschna pentacantha                                       +</td>
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<td>43570 Neoplea sp                                                    +</td>
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<td>51300 Neureclipsis sp                                               +</td>
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<tr>
<td>52200 Cheumatopsyche sp                                             +</td>
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<tr>
<td>52430 Ceratopsyche morosa group                                     +</td>
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<tr>
<td>53800 Hydroptila sp                                                 +</td>
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<tr>
<td>59410 Nectopsyche diarina                                           +</td>
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<td>60900 Peltodytes sp                                                 +</td>
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<tr>
<td>63900 Laccophilus sp                                                +</td>
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<tr>
<td>68130 Helichus sp                                                   +</td>
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<tr>
<td>68601 Ancyronyx variegata                                           +</td>
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<tr>
<td>68708 Dubiraphia vittata group                                      +</td>
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<tr>
<td>68901 Macrnychus glabratius                                         +</td>
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<tr>
<td>69400 Stenelmis sp                                                  +</td>
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<tr>
<td>71900 Tipula sp                                                     +</td>
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<tr>
<td>74100 Simulium sp                                                   +</td>
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<td>80310 Cardiocladius obscurus                                        +</td>
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<tr>
<td>80370 Corynoneura lobata                                            +</td>
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<tr>
<td>80410 Cricotopus (C.) sp                                            +</td>
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<td>80420 Cricotopus (C.) bicinctus                                     +</td>
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<td>83040 Dicrotendipes neomodestus                                    +</td>
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<td>63900 Laccophilus sp</td>
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<td>68601 Ancyronyx variegata</td>
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<td>Gyralulus (Torquis) parvus</td>
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<td>Chironomus (C.) decorus group</td>
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<td>Dicrotendipes neomodestus</td>
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<td>Paratendipes albimanus or P. duplicatus</td>
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<td>Phaenopsectra obediens group</td>
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<td>65800 Berosus sp</td>
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<td>84210 Paratendipes albinanus or P. duplicatus</td>
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<td>11651 Procloeon sp (w/o hindwing pads)</td>
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<td>78655 Procladius (Holotanypus) sp</td>
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<td>81930 Chironomus (C.) decorus group</td>
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<td>84470 Polypedilum (P.) illinoense</td>
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<tr>
<td>85230 Cladotanytarsus mancus group</td>
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<td>05900 Lirceus sp</td>
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<td>08700 Crangonyx sp</td>
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<td>11650 Procloeon sp (w/ hindwing pads)</td>
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<td>81231 Nanocladius (N.) crassicornus or N. (N.) &quot;rectinervis&quot;</td>
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<td>Polypedilum (P.) illinoense</td>
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<td>Polypedilum (Tripodura) halterale group</td>
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<tr>
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</tr>
<tr>
<td>Paratanytarsus sp</td>
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</tr>
<tr>
<td>Tanytarsus glabrescens group sp 7</td>
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</tr>
<tr>
<td>Physella sp</td>
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</tr>
<tr>
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</tr>
<tr>
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<td>Taxa</td>
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</tr>
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</tr>
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</tr>
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</tr>
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<tr>
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<tr>
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<tr>
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<tr>
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<td>16+</td>
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<tr>
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</tr>
<tr>
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<tr>
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</tr>
<tr>
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<tr>
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<tr>
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<td>Quant/Qual</td>
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<tr>
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<tr>
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<tr>
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<td>66500 Enochrus sp</td>
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<tr>
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<tr>
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<td>17200 Caenis sp</td>
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<td>2+</td>
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<td>84315 Phaenopsectra flavipes</td>
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<td>84450 Polypedilum (Uresipedia) flavum</td>
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<tr>
<td>84460 Polypedilum (P.) fallax group</td>
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<tr>
<td>84470 Polypedilum (P.) illinoense</td>
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<tr>
<td>84750 Stictochironomus sp</td>
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</tr>
<tr>
<td>85500 Paratanytarsus sp</td>
<td>253+</td>
</tr>
<tr>
<td>85625 Rheotanytarsus sp</td>
<td>190+</td>
</tr>
<tr>
<td>85821 Tanytarsus glabrescens group sp 7</td>
<td>316+</td>
</tr>
<tr>
<td>87540 Hemerodromia sp</td>
<td>17+</td>
</tr>
<tr>
<td>96900 Ferrissia sp</td>
<td>37+</td>
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<tr>
<td>99160 Anodontoides ferussacianus</td>
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</tr>
<tr>
<td>99180 Strophitus undulatus undulatus</td>
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<td>Taxa</td>
<td>Quant/Qual</td>
</tr>
<tr>
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</tr>
<tr>
<td>01320 Hydra sp</td>
<td>448 85720</td>
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<tr>
<td>01801 Turbellaria</td>
<td>392+ 85800</td>
</tr>
<tr>
<td>03600 Oligochaeta</td>
<td>64+ 85821</td>
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<tr>
<td>05900 Lirceus sp</td>
<td>1 85840</td>
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<tr>
<td>08250 Orconectes (Procericambarus) rusticus</td>
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<tr>
<td>08601 Hydrachnidia</td>
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<tr>
<td>11020 Acerpenna pygmaea</td>
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<tr>
<td>11130 Baetis intercalaris</td>
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</tr>
<tr>
<td>11400 Clinostomum sp</td>
<td>+ 99240</td>
</tr>
<tr>
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<td>13400 Stenacron sp</td>
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</tr>
<tr>
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<tr>
<td>16700 Tricorythodes sp</td>
<td>51+</td>
</tr>
<tr>
<td>17200 Caenis sp</td>
<td>1+</td>
</tr>
<tr>
<td>21300 Hetaerina sp</td>
<td>51+</td>
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<tr>
<td>22001 Coenagrionidae</td>
<td>+</td>
</tr>
<tr>
<td>22300 Argia sp</td>
<td>53+</td>
</tr>
<tr>
<td>24900 Gomphus sp</td>
<td>+</td>
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<tr>
<td>44501 Corixidae</td>
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<tr>
<td>52200 Cheumatopsyche sp</td>
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</tr>
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<td>992+</td>
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</tr>
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</tr>
<tr>
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<td>1+</td>
</tr>
<tr>
<td>67700 Paracymus sp</td>
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<tr>
<td>68901 Macrorychus glabratus</td>
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</tr>
<tr>
<td>69400 Stenelmis sp</td>
<td>2+</td>
</tr>
<tr>
<td>71900 Tipula sp</td>
<td>3+</td>
</tr>
<tr>
<td>72700 Anopheles sp</td>
<td>+</td>
</tr>
<tr>
<td>77120 Ablabesmyia mallochi</td>
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</tr>
<tr>
<td>77750 Thienemannimyia sp</td>
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<tr>
<td>78140 Labrundinia pilosella</td>
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<td>80370 Corynoneura lobata</td>
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<tr>
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<tr>
<td>82822 Cryptochironomus eminencia</td>
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<tr>
<td>83040 Dicrotendipes neomodestus</td>
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<tr>
<td>83820 Microtendipes &quot;caelum&quot; (sensu Simpson &amp; Bode, 1980)</td>
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<tr>
<td>84450 Polypedilum (Uresipedium) flavum</td>
<td>4,907+</td>
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<tr>
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<tr>
<td>84520 Polypedilum (Tripodura) halterale group</td>
<td>+</td>
</tr>
<tr>
<td>84540 Polypedilum (Tripodura) scalaenum group</td>
<td>+</td>
</tr>
<tr>
<td>84612 Saetheria tylus</td>
<td>+</td>
</tr>
<tr>
<td>84750 Stictochironomus sp</td>
<td>+</td>
</tr>
<tr>
<td>85230 Cladotanytarsus mancus group</td>
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<td>85264 Cladotanytarsus vanderwulpi group sp 4</td>
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<td>Quant/Qual</td>
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<td>01320 Hydra sp</td>
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<tr>
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<td>+ 85800 Tanytarsus sp</td>
</tr>
<tr>
<td>07875 Cambarus (Tubercambarus)polychromatus</td>
<td>+ 85821 Tanytarsus glabrescens group sp 7 851</td>
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</tr>
<tr>
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<tr>
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</tr>
<tr>
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<tr>
<td>21200 Calopteryx sp</td>
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<tr>
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<tr>
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</tr>
<tr>
<td>22300 Arigma sp</td>
<td>26+</td>
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<td>24900 Gomphus sp</td>
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<tr>
<td>27307 Epiphanus (Epiphanus) princeps</td>
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<tr>
<td>51600 Polycenotopus sp</td>
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<tr>
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<tr>
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<tr>
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<tr>
<td>59300 Mystacides sp</td>
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<tr>
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<tr>
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<tr>
<td>65800 Berosus sp</td>
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<tr>
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</tr>
<tr>
<td>68901 Macrourus glabatus</td>
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<tr>
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<td>57+</td>
</tr>
<tr>
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<td>74100 Simulium sp</td>
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<tr>
<td>77130 Ablabesmyia hamphe group</td>
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<td>84210 Paratendipes albinanus or P. duplicatus</td>
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<tr>
<td>84300 Phaenopsectra obediens group</td>
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<tr>
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<tr>
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<tr>
<td>84470 Polypedilum (P.) illinoense</td>
<td>+</td>
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<tr>
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<tr>
<td>Taxa</td>
<td>Quant/Qual</td>
</tr>
<tr>
<td>-------------------------------------------</td>
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</tr>
<tr>
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<td>+</td>
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<tr>
<td>02600 Nematomorpha</td>
<td>+</td>
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<tr>
<td>04685 Placobdella ornata</td>
<td>+</td>
</tr>
<tr>
<td>04935 Erpobdella punctata punctata</td>
<td>+</td>
</tr>
<tr>
<td>05900 Lirceus sp</td>
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</tr>
<tr>
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<tr>
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<tr>
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<td>+</td>
</tr>
<tr>
<td>13400 Stenacron sp</td>
<td>+</td>
</tr>
<tr>
<td>13521 Stenonema femoratum</td>
<td>+</td>
</tr>
<tr>
<td>17200 Caenis sp</td>
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<tr>
<td>22001 Coenagrionidae</td>
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</tr>
<tr>
<td>22300 Argia sp</td>
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</tr>
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<tr>
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</tr>
<tr>
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<tr>
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<td>+</td>
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<tr>
<td>68708 Dubraphia vittata group</td>
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<tr>
<td>69400 Stenelmis sp</td>
<td>+</td>
</tr>
<tr>
<td>69420 Stenelmis sexlineata</td>
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<td>71900 Tipula sp</td>
<td>+</td>
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<tr>
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<tr>
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<tr>
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<tr>
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<tr>
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<tr>
<td>85625 Rheotanytarsus sp</td>
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<tr>
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</tr>
<tr>
<td>95100 Physella sp</td>
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<tr>
<td>96280 Planorbella (Pierosoma) trivolvis</td>
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<tr>
<td>98600 Sphaerium sp</td>
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<tr>
<td>Taxa</td>
<td>Quant/Qual</td>
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</tr>
<tr>
<td>01320 Hydra sp</td>
<td>17+</td>
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<tr>
<td>01801 Turbellaria</td>
<td>65+</td>
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<tr>
<td>03600 Oligochaeta</td>
<td>1+</td>
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<tr>
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<td>+</td>
</tr>
<tr>
<td>08601 Hydrachnidia</td>
<td>1+</td>
</tr>
<tr>
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<td>171+</td>
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<tr>
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<td>11130 Baetis intercalaris</td>
<td>37+</td>
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<tr>
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<tr>
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<tr>
<td>22300 Hetaerina sp</td>
<td>1+</td>
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<tr>
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<td>1+</td>
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<tr>
<td>65800 Berosus sp</td>
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</tr>
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<td>66500 Enochus sp</td>
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<tr>
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<td>+</td>
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<tr>
<td>68300 Cyphon sp</td>
<td>+</td>
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<tr>
<td>69400 Stenelmis sp</td>
<td>16+</td>
</tr>
<tr>
<td>71900 Tipula sp</td>
<td>+</td>
</tr>
<tr>
<td>72700 Anopheles sp</td>
<td>+</td>
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<tr>
<td>77120 Ablabesmyia mallochi</td>
<td>78+</td>
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<tr>
<td>77500 Conchapelopia sp</td>
<td>117+</td>
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<td>78140 Labrundinia pilosella</td>
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<tr>
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<tr>
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<td>80420 Cricotopus (C.) bicinctus</td>
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<tr>
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<td>39+</td>
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<tr>
<td>83040 Dicrotendipes neomodestus</td>
<td>39+</td>
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<td>84116 Paracaedopelma nereis</td>
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<td>84210 Paratendipes albimans or P. duplicatus</td>
<td>78</td>
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<tr>
<td>84315 Phaenopsectra flavipes</td>
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<td>84450 Polypedilum (Uresipedilum) flavum</td>
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<td>85800 Tanytarsus sp</td>
<td>195+</td>
</tr>
<tr>
<td>85821 Tanytarsus glabrescens group sp 7</td>
<td>781+</td>
</tr>
<tr>
<td>85840 Tanytarsus sepp</td>
<td>78+</td>
</tr>
<tr>
<td>95100 Physella sp</td>
<td>10+</td>
</tr>
<tr>
<td>96284 Planorbella (Pierosoma) pilbsryi</td>
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</tr>
<tr>
<td>96900 Ferrissia sp</td>
<td>120+</td>
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<tr>
<td>98600 Sphaerium sp</td>
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<tr>
<td>Taxa</td>
<td>Quant/Qual</td>
</tr>
<tr>
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<tr>
<td>Hydra sp</td>
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<tr>
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<tr>
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<td>Helodella papillata</td>
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<tr>
<td>Erpobdella microstoma</td>
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<tr>
<td>Lirceus sp</td>
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<tr>
<td>Acerpenna pygmaea</td>
<td>59+</td>
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<tr>
<td>Baetis flavistriga</td>
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<tr>
<td>Baetis intercalaris</td>
<td>8+</td>
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<tr>
<td>Stenacron sp</td>
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<td>Belostoma sp</td>
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<td>Cheumatopsyche sp</td>
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<tr>
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<tr>
<td>Hydroptila sp</td>
<td>229+</td>
</tr>
<tr>
<td>Berosus sp</td>
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<tr>
<td>Tropisternus sp</td>
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<tr>
<td>Dubraphia vittata group</td>
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<td>Limonia sp</td>
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<td>Tipula sp</td>
<td>11+</td>
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<td>Anopheles sp</td>
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<tr>
<td>Ablabesmyia mallochi</td>
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<tr>
<td>Conchapelopia sp</td>
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<tr>
<td>Thienemannimyia sp</td>
<td>231+</td>
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<td>Corynoneura lobata</td>
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<tr>
<td>Nano cladius (N.) spinipennis</td>
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<td>Dicrotendipes sp</td>
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<td>Quant/Qual</td>
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<tr>
<td>08250 Orconectes (Procericambarus) rusticus</td>
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<td>+</td>
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<tr>
<td>11130 Baetis intercalaris</td>
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<td>13521 Stenonema femoratum</td>
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<tr>
<td>22001 Coenagrionidae</td>
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<td>52430 Ceratopsyche morosa group</td>
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<td>52530 Hydropsyche depravata group</td>
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<tr>
<td>60900 Peltodytes sp</td>
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<tr>
<td>69400 Stenelmis sp</td>
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<tr>
<td>71900 Tipula sp</td>
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<tr>
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<tr>
<td>11651 Procloeon sp (w/o hindwing pads)</td>
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<tr>
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<tr>
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<td>Taxa</td>
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<td>Crangonyx sp</td>
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<tr>
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<tr>
<td>Procloeon sp (w/o hindwing pads)</td>
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<tr>
<td>Stenacron sp</td>
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<tr>
<td>Stenonema femoratum</td>
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<tr>
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<tr>
<td>Dicrotendipes neomodestus</td>
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<tr>
<td>Microtendipes &quot;caelum&quot; (sensu Simpson &amp; Bode, 1980)</td>
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<tr>
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<tr>
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<tr>
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<td>Fossaria sp</td>
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<td>Taxa</td>
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<tr>
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<td>Baetis intercalaris</td>
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<td>Hydropsyche depravata group</td>
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<td>Anopheles sp</td>
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<tr>
<td>Simulium sp</td>
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<td>Nilotanypus fimbriatus</td>
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<td>Corynoneura floridensis</td>
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<td>Corynoneura lobata</td>
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<td>Cricotopus sp</td>
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<td>Polypedilum (Uresipedilum) flavum</td>
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<td>Paratanytarsus sp</td>
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<tr>
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<td>Leucrocuta sp</td>
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<td>Argia sp</td>
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<td>Cheumatopsyche sp</td>
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<td>Peltodytes sp</td>
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<td>Copelatus sp</td>
<td>+</td>
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<td>Stenelmis sp</td>
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<td>Simulium sp</td>
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<td>Paratendipes albimanus or P. duplicatus</td>
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<tr>
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Rivercode: 04-042-000  
River Mile: 2.30  
0:00 9/9/15  

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### SNOOKS RUN AT THE BEND @ SLOUGH RD.

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**Rivercode:** 04-049-000  
**River Mile:** 0.63  
**Date:** 9/22/15

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**Narrative:**

0:00 9/22/15

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- **Erpobdella microstoma**: +
- **Lirceus sp**: +
- **Cranonyx sp**: +
- **Callibaetis sp**: +
- **Coenagrionidae**: +
- **Sigara sp**: +
- **Cheumatopsyche sp**: +
- **Peltodytes sp**: +
- **Stenelmis sp**: +
- **Tipula sp**: +
- **Dixella sp**: +
- **Anopheles sp**: +
- **Conchapelopia sp**: +
- **Procladius (Holotanypus) sp**: +
- **Chironomus (C.) decorus group**: +
- **Kiefferulus sp**: +
- **Paratendipes albimanus or P. duplicatus**: +
- **Polyplegium (P.) illinoense**: +
- **Tanytarsus sp**: +
- **Stagnicola sp**: +
- **Physella sp**: +

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**River Mile:** 0.78  
**Date:** 0:00 9/22/15
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### M. FK. GORDON CREEK @ LAKE RD.

**Station:** P06S18  
**Rivercode:** 04-055-000  
**River Mile:** 3.80  
**0:00  8/30/16**

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River Mile: 0.10
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S. BR. MARIE DELARME CREEK N OF ANTWERP @ CO. RD. 45  
0:00 8/6/15

Rivercode: 04-060-000  
River Mile: 1.00

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<tr>
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Appendix B

Macroinvertebrate ICI and Metric Results
### Invertebrate Community Index metrics for stations sampled in the Maumee River Tributaries survey, 2015.

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<th>Percent Composition</th>
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### Invertebrate Community Index metrics for stations sampled in the Maumee River Tributaries survey, 2015.

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Appendix C

Fish Species and Abundance Results
**Species Name** | **IBI** | **Feed Guild** | **Breed Guild** | **Tol** | **# of Fish** | **Relative Number** | **% by Number** | **Relative Weight** | **% by Weight** | **Ave Weight (gm)**
---|---|---|---|---|---|---|---|---|---|---
White Sucker | O | S | T | 2 | 3.24 | 0.30 | 0.06 | 0.78 | 17.00
Goldfish | O | M | T | 4 | 6.49 | 0.59 | 0.53 | 7.45 | 81.00
Western Blacknose Dace | G | S | T | 1 | 1.62 | 0.15 | 0.01 | 0.09 | 4.00
Creek Chub | G | N | T | 79 | 128.11 | 11.69 | 1.19 | 16.84 | 9.27
Suckermouth Minnow | I | S | N | 2 | 3.24 | 0.30 | 0.01 | 0.14 | 3.00
Redfin Shiner | I | N | N | 1 | 1.62 | 0.15 | 0.00 | 0.05 | 2.00
Common Shiner | I | S | N | 53 | 85.95 | 7.84 | 0.33 | 4.74 | 3.89
Spotfin Minnow | I | M | N | 15 | 24.32 | 2.22 | 0.06 | 0.87 | 2.53
Sand Shiner | I | M | M | 13 | 21.08 | 1.92 | 0.03 | 0.45 | 1.50
Fathead Minnow | O | C | T | 13 | 21.08 | 1.92 | 0.04 | 0.60 | 2.00
Bluntnose Minnow | O | C | T | 106 | 171.89 | 15.68 | 0.22 | 3.10 | 1.27
Central Stoneroller | H | N | N | 35 | 56.76 | 5.18 | 0.47 | 6.62 | 8.23
Yellow Bullhead | I | C | T | 6 | 9.73 | 0.89 | 0.33 | 4.69 | 34.00
Blackstripe Topminnow | I | M | N | 9 | 14.59 | 1.33 | 0.02 | 0.31 | 1.50
Green Sunfish | I | C | T | 262 | 424.86 | 38.76 | 3.48 | 49.40 | 8.20
Bluegill Sunfish | I | C | P | 2 | 3.24 | 0.30 | 0.07 | 1.06 | 23.00
Hybrid x Sunfish | N | 4 | 6.49 | 0.59 | 0.08 | 1.10 | 12.00
Johnny Darter | I | C | N | 32 | 51.89 | 4.73 | 0.07 | 1.05 | 1.43
Orangethroat Darter | I | S | N | 37 | 60.00 | 5.47 | 0.05 | 0.67 | 0.79

**Data Totals:**
676 | 1096.22 | 7.05

**Number of Species:** 19

**Number of Hybrids:** 1
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<th>Feed Guild</th>
<th>Breed Guild</th>
<th>Tol</th>
<th># of Fish</th>
<th>Relative Number</th>
<th>% by Number</th>
<th>Relative Weight</th>
<th>% by Weight</th>
<th>Ave Weight (gm)</th>
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**Sample Date:** 2015-08-31  
**Location:** TONTOGANY CREEK N OF TONTOGANY @ ROBINSON RD.  
**Depth:** N  
**Drainage:** 39.4 sq mi  
**Basin:** Maumee River  
**Data Source:**  
**Sampler Type:** D

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**Sample Date:** 2015-08-06  
**River Mile:** 9.0  
**Drainage:** 10.0 sq mi  
**Basin:** Maumee River  
**Time Fished:** 900 sec  
**Depth:** N  
**Dist Fished:** 0.15 km  
**Flow:** N  
**Sampler Type:** E

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- **Stream:** BIG CREEK
- **Sample Date:** 2015-07-28
- **River Mile:** 1.3
- **Location:** BIG CREEK N OF MCCLURE @ TWP. RD. Q
- **Drainage:** 20.7 sq mi
- **Basin:** Maumee River
- **Flow:** N
- **Time Fished:** 1800 sec
- **Dist Fished:** 0.16 km

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- **Depth:** Data Source: E

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### Drainage: 23.9 sq mi
### Basin: Maumee River
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### Time Fished: 2700 sec
### Dist Fished: 0.16 km
### Data Source: E
### Sampler Type: E

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**Sample Date:** 2015-09-03  
**Drainage:** 65.0 sq mi  
**Basin:** Maumee River  
**Time Fished:** 2700 sec  
**Dist Fished:** 0.17 km  
**Location:** S. TURKEYFOOT CREEK AT MALINTA @ CO. RD. L

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Drainage: 8.3 sq mi
Basin: Maumee River
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**Dist Fished:** 0.15 km  
**Sample Date:** 2015-07-23  
**Location:** PLATTER CREEK @ FARMER MARK RD.  
**Drainage:** 11.9 sq mi  
**Basin:** Maumee River  
**Flow:** N  
**Sampler Type:** E

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**Sample Date:** 2015-07-23  
**River Mile:** 1.3  
**Location:** N. FORK GORDON CREEK @ ROSEDALE RD.  
**Drainage:** 11.1 sq mi  
**Basin:** Maumee River  
**Depth:**  
**Flow:** N  
**Dist Fished:** 0.2 km  
**Sampler Type:** E

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Appendix D

Fish IBI Metric and MIwb Results
Appendix x. Index of Biotic Integrity component metrics and scores for headwater locations sampled during the 2015 Maumee River Tributaries survey.

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<th>Drain Area</th>
<th>Total Species</th>
<th>Minnow Species</th>
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* Less than 200 total individuals in the sample, values appear as superscripts.

Project Number: 794 Maumee River Basin Select Tributaries 2015 TMDL
Appendix x. Index of Biotic Integrity component metrics and scores for headwater locations sampled during the 2015 Maumee River Tributaries survey.

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* Less than 200 total individuals in the sample, values appear as superscripts
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<th>Minnow Species</th>
<th>Headwater Species</th>
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* Less than 200 total individuals in the sample, values appear as superscripts
Appendix x. Index of Biotic Integrity component metrics and scores for headwater locations sampled during the 2015 Maumee River Tributaries survey.

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<tr>
<th>RM</th>
<th>Type</th>
<th>Date</th>
<th>Drain Area</th>
<th>Total Species</th>
<th>Minnow Species</th>
<th>Headwater Species</th>
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<th>Darter Sculpin Species</th>
<th>Simple Lithophils</th>
<th>Tolerant Fishes</th>
<th>Omni-vores</th>
<th>Pioneering Fishes</th>
<th>Insect-ivores</th>
<th>DELT Anomalies</th>
<th>Rel. No. * minus tolerants</th>
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* Less than 200 total individuals in the sample, values appear as superscripts.
Appendix x. Index of Biotic Integrity component metrics and scores for headwater locations sampled during the 2015 Maumee River Tributaries survey.

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<tr>
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<th>Tolerant Fishes</th>
<th>Omni-vores</th>
<th>Pioneering Fishes</th>
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* Less than 200 total individuals in the sample, values appear as superscripts
### Appendix x. Index of Biotic Integrity component metrics and scores for wadeable locations sampled from Maumee River tributaries, 2015.

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<tr>
<th>RM</th>
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### Appendix x. Index of Biotic Integrity component metrics and scores for wadeable locations sampled from Maumee River tributaries, 2015.

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### Index of Biotic Integrity component metrics and scores for wadeable locations sampled from Maumee River tributaries, 2015.

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<th>Rel. No. minus tolerant</th>
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**Project Number:** Maumee River Basin Select Tributaries 2015 TMDL
### Appendix x. Index of Biotic Integrity component metrics and scores for wadeable locations sampled from Maumee River tributaries, 2015.

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<th>Date</th>
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<th>Darter Species</th>
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* Less than 200 total individuals in the sample, values appear as superscripts.
Appendix E

Water Quality Sonde Monitoring – Hourly Plots of Field Parameters
Appendix X: Sonde data summary plots

Figures 1-31 show the August 4-8, 2015 survey sonde data. Figures 34-47 show the August 12-17, 2015 survey sonde data. Figures 48-69 show the September 9-11, 2015 survey sonde data. Figures 70-81 show the June 14-15, 2016 survey sonde data. On all figures in this appendix the values for first three parameters, temperature, pH and dissolved oxygen (D.O.), are shown on the left side of the plot. The values for specific conductance are shown on the right.

Figure 1: Plot of hourly data collected with a water quality sonde on Zuber Cutoff (RM 1.2; STORET: P06K14). Temperature (°C), D.O. (mg/L), pH (S.U.) and specific conductance (µS/cm) are included. The data was collected from 8/4-6/2015.

Figure 2: Plot of hourly data collected with a water quality sonde on Marie DeLarme (RM 0.5; STORET: P06K24). Temperature (°C), D.O. (mg/L), pH (S.U.) and specific conductance (µS/cm) are included. The data was collected from 8/4-6/2015.
Figure 3: Plot of hourly data collected with a water quality sonde on Gordon Creek (RM 1.12; STORET: P06S04). Temperature (°C), D.O. (mg/L), pH (S.U.) and specific conductance (µS/cm) are included. The data was collected from 8/4-6/2015.

Figure 4: Plot of hourly data collected with a water quality sonde on Platter Creek (RM 1.7; STORET: 303010). Temperature (°C), D.O. (mg/L), pH (S.U.) and specific conductance (µS/cm) are included. The data was collected from 8/4-6/2015.
Figure 5: Plot of hourly data collected with a water quality sonde on Benien Creek (RM 2.3; STORET: P09K18). Temperature (°C), D.O. (mg/L), pH (S.U.) and specific conductance (µS/cm) are included. The data was collected from 8/4-5/2015.

Figure 6: Plot of hourly data collected with a water quality sonde on Garrett Creek (RM 0.7; STORET: P09K17). Temperature (°C), D.O. (mg/L), pH (S.U.) and specific conductance (µS/cm) are included. The data was collected from 8/4-6/2015.
Figure 7: Plot of hourly data collected with a water quality sonde on Van Hying Creek (RM 0.75; STORET: 302983). Temperature (°C), D.O. (mg/L), pH (S.U.) and specific conductance (µS/cm) are included. The data was collected from 8/4-6/2015.

Figure 8: Plot of hourly data collected with a water quality sonde on South Turkeyfoot Creek (RM 19.75; STORET: P09526). Temperature (°C), D.O. (mg/L), pH (S.U.) and specific conductance (µS/cm) are included. The data was collected from 8/4-6/2015.
Figure 9: Plot of hourly data collected with a water quality sonde on South Turkeyfoot Creek (RM 7.9; STORET: 302836). Temperature (°C), D.O. (mg/L), pH (S.U.) and specific conductance (µS/cm) are included. The data was collected from 8/4-6/2015.

Figure 10: Plot of hourly data collected with a water quality sonde on West Creek (RM 1; STORET: P10K07). Temperature (°C), D.O. (mg/L), pH (S.U.) and specific conductance (µS/cm) are included. The data was collected from 8/4-6/2015.
Figure 11: Plot of hourly data collected with a water quality sonde on Lost Creek (RM 1.3; STORET: P09S09). Temperature (°C), D.O. (mg/L), pH (S.U.) and specific conductance (µS/cm) are included. The data was collected from 8/4-6/2015.

Figure 12: Plot of hourly data collected with a water quality sonde on School Creek (RM 0.9; STORET: 302994). Temperature (°C), D.O. (mg/L), pH (S.U.) and specific conductance (µS/cm) are included. The data was collected from 8/4-6/2015.
Figure 13: Plot of hourly data collected with a water quality sonde on Little Turkeyfoot Creek (RM 0.48; STORET: 302843). Temperature (°C), D.O. (mg/L), pH (S.U.) and specific conductance (µS/cm) are included. The data was collected from 8/4-6/2015.

Figure 14: Plot of hourly data collected with a water quality sonde on North Turkeyfoot Creek (RM 17.85; STORET: P09504). Temperature (°C), D.O. (mg/L), pH (S.U.) and specific conductance (µS/cm) are included. The data was collected from 8/4-6/2015.
Figure 15: Plot of hourly data collected with a water quality sonde on North Turkeyfoot Creek (RM 3.4; STORET: P09S01). Temperature (°C), D.O. (mg/L), pH (S.U.) and specific conductance (µS/cm) are included. The data was collected from 8/4-6/2015.

Figure 16: Plot of hourly data collected with a water quality sonde on Konzen Ditch (RM 0.65; STORET: P09K14). Temperature (°C), D.O. (mg/L), pH (S.U.) and specific conductance (µS/cm) are included. The data was collected from 8/4-6/2015.
Figure 17: Plot of hourly data collected with a water quality sonde on Dry Creek (RM 1.6; STORET: 302990). Temperature (°C), D.O. (mg/L), pH (S.U.) and specific conductance (µS/cm) are included. The data was collected from 8/4-6/2015.

Figure 18: Plot of hourly data collected with a water quality sonde on Bad Creek (RM 17.51; STORET: P11W22). Temperature (°C), D.O. (mg/L), pH (S.U.) and specific conductance (µS/cm) are included. The data was collected from 8/4-6/2015.
Figure 19: Plot of hourly data collected with a water quality sonde on Bad Creek (RM 10.46; STORET: P11S05). Temperature (°C), D.O. (mg/L), pH (S.U.) and specific conductance (µS/cm) are included. The data was collected from 8/4-6/2015.

Figure 20: Plot of hourly data collected with a water quality sonde on Bad Creek (RM 2.47; STORET: P11S04). Temperature (°C), D.O. (mg/L), pH (S.U.) and specific conductance (µS/cm) are included. The data was collected from 8/4-6/2015.
Figure 21: Plot of hourly data collected with a water quality sonde on Big Creek (RM 1.3; STORET: P09K06). Temperature (°C), D.O. (mg/L), pH (S.U.) and specific conductance (µS/cm) are included. The data was collected from 8/4-6/2015.

Figure 22: Plot of hourly data collected with a water quality sonde on Beaver Creek (RM 8.3; STORET: P10K03). Temperature (°C), D.O. (mg/L), pH (S.U.) and specific conductance (µS/cm) are included. The data was collected from 8/4-6/2015.
Figure 23: Plot of hourly data collected with a water quality sonde on Beaver Creek (RM 2.73; STORET: P10K02). Temperature (°C), D.O. (mg/L), pH (S.U.) and specific conductance (µS/cm) are included. The data was collected from 8/4-6/2015.

Figure 24: Plot of hourly data collected with a water quality sonde on Hammer Creek (RM 1.34; STORET: 303000). Temperature (°C), D.O. (mg/L), pH (S.U.) and specific conductance (µS/cm) are included. The data was collected from 8/4-6/2015.
Figure 25: Plot of hourly data collected with a water quality sonde on Jackson Cutoff Ditch (RM 1.15; STORET: 510040). Temperature (°C), D.O. (mg/L), pH (S.U.) and specific conductance (µS/cm) are included. The data was collected from 8/4-6/2015.

Figure 26: Plot of hourly data collected with a water quality sonde on Little Yellow Creek (RM 0.9; STORET: 500700). Temperature (°C), D.O. (mg/L), pH (S.U.) and specific conductance (µS/cm) are included. The data was collected from 8/4-6/2015.
Figure 27: Plot of hourly data collected with a water quality sonde on West Creek (trib to Yellow Ck) (RM 0.1; STORET: 302840). Temperature (°C), D.O. (mg/L), pH (S.U.) and specific conductance (µS/cm) are included. The data was collected from 8/4-6/2015.

Figure 28: Plot of hourly data collected with a water quality sonde on Brush Creek (RM 0.58; STORET: P10P06). Temperature (°C), D.O. (mg/L), pH (S.U.) and specific conductance (µS/cm) are included. The data was collected from 8/4-6/2015.
Figure 29: Plot of hourly data collected with a water quality sonde on Tontogany Creek (RM 1.6; STORET: P10K01). Temperature (°C), D.O. (mg/L), pH (S.U.) and specific conductance (µS/cm) are included. The data was collected from 8/4-6/2015.

Figure 30: Plot of hourly data collected with a water quality sonde on West Branch Tontogany Creek (RM 3.42; STORET: P10P13). Temperature (°C), D.O. (mg/L), pH (S.U.) and specific conductance (µS/cm) are included. The data was collected from 8/4-6/2015.
Figure 31: Plot of hourly data collected with a water quality sonde on Liberty High Road Ditch (RM 0.05; STORET: 303008). Temperature (°C), D.O. (mg/L), pH (S.U.) and specific conductance (µS/cm) are included. The data was collected from 8/4-6/2015.

Figure 32: Plot of hourly data collected with a water quality sonde on Benien Creek (RM 2.3; STORET: P09K18). Temperature (°C), D.O. (mg/L), pH (S.U.) and specific conductance (µS/cm) are included. The data was collected from 8/12-17/2015.
Figure 33: Plot of hourly data collected with a water quality sonde on Garrett Creek (RM 0.7; STORET: P09K17). Temperature (°C), D.O. (mg/L), pH (S.U.) and specific conductance (µS/cm) are included. The data was collected from 8/12-17/2015.

Figure 34: Plot of hourly data collected with a water quality sonde on Van Hyning Creek (RM 0.75; STORET: 302983). Temperature (°C), D.O. (mg/L), pH (S.U.) and specific conductance (µS/cm) are included. The data was collected from 8/12-17/2015.
Figure 35: Plot of hourly data collected with a water quality sonde on South Turkeyfoot Creek (RM 7.9; STORET: 302836). Temperature (°C), D.O. (mg/L), pH (S.U.) and specific conductance (µS/cm) are included. The data was collected from 8/12-17/2015.

Figure 36: Plot of hourly data collected with a water quality sonde on School Creek (RM 0.9; STORET: 302994). Temperature (°C), D.O. (mg/L), pH (S.U.) and specific conductance (µS/cm) are included. The data was collected from 8/12-17/2015.
Figure 37: Plot of hourly data collected with a water quality sonde on Little Turkeyfoot Creek (RM 0.48; STORET: 302843). Temperature (°C), D.O. (mg/L), pH (S.U.) and specific conductance (µS/cm) are included. The data was collected from 8/12-17/2015.

Figure 38: Plot of hourly data collected with a water quality sonde on Dry Creek (RM 1.6; STORET: 302990). Temperature (°C), D.O. (mg/L), pH (S.U.) and specific conductance (µS/cm) are included. The data was collected from 8/12-17/2015.
Figure 39: Plot of hourly data collected with a water quality sonde on Bad Creek (RM 17.51; STORET: P11W22). Temperature (°C), D.O. (mg/L), pH (S.U.) and specific conductance (µS/cm) are included. The data was collected from 8/12-17/2015.

Figure 40: Plot of hourly data collected with a water quality sonde on Bad Creek (RM 10.46; STORET: P11S05). Temperature (°C), D.O. (mg/L), pH (S.U.) and specific conductance (µS/cm) are included. The data was collected from 8/12-17/2015.
Figure 41: Plot of hourly data collected with a water quality sonde on Bad Creek (RM 2.47; STORET: P11S04). Temperature (°C), D.O. (mg/L), pH (S.U.) and specific conductance (µS/cm) are included. The data was collected from 8/12-17/2015.

Figure 42: Plot of hourly data collected with a water quality sonde on Big Creek (RM 1.3; STORET: P09K06). Temperature (°C), D.O. (mg/L), pH (S.U.) and specific conductance (µS/cm) are included. The data was collected from 8/12-17/2015.
Figure 43: Plot of hourly data collected with a water quality sonde on Beaver Creek (RM 8.3; STORET: P10K03). Temperature (°C), D.O. (mg/L), pH (S.U.) and specific conductance (µS/cm) are included. The data was collected from 8/12-17/2015.

Figure 44: Plot of hourly data collected with a water quality sonde on Beaver Creek (RM 2.73; STORET: P10K02). Temperature (°C), D.O. (mg/L), pH (S.U.) and specific conductance (µS/cm) are included. The data was collected from 8/12-17/2015.
Figure 45: Plot of hourly data collected with a water quality sonde on Hammer Creek (RM 1.34; STORET: 303000). Temperature (°C), D.O. (mg/L), pH (S.U.) and specific conductance (µS/cm) are included. The data was collected from 8/12-17/2015.

Figure 46: Plot of hourly data collected with a water quality sonde on Jackson Cutoff Ditch (RM 1.15; STORET: 510040). Temperature (°C), D.O. (mg/L), pH (S.U.) and specific conductance (µS/cm) are included. The data was collected from 8/12-17/2015.
Figure 47: Plot of hourly data collected with a water quality sonde on West Creek (trib to Yellow Ck) (RM 0.1; STORET: 302840). Temperature (°C), D.O. (mg/L), pH (S.U.) and specific conductance (µS/cm) are included. The data was collected from 8/12-17/2015.

Figure 48: Plot of hourly data collected with a water quality sonde on South Creek (RM 2.08; STORET: 302970). Temperature (°C), D.O. (mg/L), pH (S.U.) and specific conductance (µS/cm) are included. The data was collected from 9/9-11/2015.
Figure 49: Plot of hourly data collected with a water quality sonde on Marie DeLarme (RM 0.5; STORET: P06K24). Temperature (°C), D.O. (mg/L), pH (S.U.) and specific conductance (µS/cm) are included. The data was collected from 9/9-11/2015.

Figure 50: Plot of hourly data collected with a water quality sonde on Platter Creek (RM 1.7; STORET: 303010). Temperature (°C), D.O. (mg/L), pH (S.U.) and specific conductance (µS/cm) are included. The data was collected from 9/9-11/2015.
Figure 51: Plot of hourly data collected with a water quality sonde on Benien Creek (RM 2.3; STORET: P09K18). Temperature (°C), D.O. (mg/L), pH (S.U.) and specific conductance (µS/cm) are included. The data was collected from 9/9-11/2015.

Figure 52: Plot of hourly data collected with a water quality sonde on Garrett Creek (RM 0.7; STORET: P09K17). Temperature (°C), D.O. (mg/L), pH (S.U.) and specific conductance (µS/cm) are included. The data was collected from 9/9-11/2015.
Figure 53: Plot of hourly data collected with a water quality sonde on Van Hyning Creek (RM 0.75; STORET: 302983). Temperature (°C), D.O. (mg/L), pH (S.U.) and specific conductance (µS/cm) are included. The data was collected from 9/9-11/2015.

Figure 54: Plot of hourly data collected with a water quality sonde on South Turkeyfoot Creek (RM 19.75; STORET: P09S26). Temperature (°C), D.O. (mg/L), pH (S.U.) and specific conductance (µS/cm) are included. The data was collected from 9/9-11/2015.
Figure 55: Plot of hourly data collected with a water quality sonde on South Turkeyfoot Creek (RM 7.9; STORET: 302836). Temperature (°C), D.O. (mg/L), pH (S.U.) and specific conductance (µS/cm) are included. The data was collected from 9/9-11/2015.

Figure 56: Plot of hourly data collected with a water quality sonde on Little Turkeyfoot Creek (RM 0.48; STORET: 302843). Temperature (°C), D.O. (mg/L), pH (S.U.) and specific conductance (µS/cm) are included. The data was collected from 9/9-11/2015.
Figure 57: Plot of hourly data collected with a water quality sonde on North Turkeyfoot Creek (RM 17.85; STORET: P09S04). Temperature (°C), D.O. (mg/L), pH (S.U.) and specific conductance (µS/cm) are included. The data was collected from 9/9-11/2015.

Figure 58: Plot of hourly data collected with a water quality sonde on North Turkeyfoot Creek (RM 3.4; STORET: P09S01). Temperature (°C), D.O. (mg/L), pH (S.U.) and specific conductance (µS/cm) are included. The data was collected from 9/9-11/2015.
Figure 59: Plot of hourly data collected with a water quality sonde on Konzen Ditch (RM 0.65; STORET: P09K14). Temperature (°C), D.O. (mg/L), pH (S.U.) and specific conductance (µS/cm) are included. The data was collected from 9/9-11/2015.

Figure 60: Plot of hourly data collected with a water quality sonde on Dry Creek (RM 1.6; STORET: 302990). Temperature (°C), D.O. (mg/L), pH (S.U.) and specific conductance (µS/cm) are included. The data was collected from 9/9-11/2015.
Figure 61: Plot of hourly data collected with a water quality sonde on Bad Creek (RM 17.51; STORET: P11W22). Temperature (°C), D.O. (mg/L), pH (S.U.) and specific conductance (µS/cm) are included. The data was collected from 9/9-11/2015.

Figure 62: Plot of hourly data collected with a water quality sonde on Bad Creek (RM 2.47; STORET: P11S04). Temperature (°C), D.O. (mg/L), pH (S.U.) and specific conductance (µS/cm) are included. The data was collected from 9/9-11/2015.
Figure 63: Plot of hourly data collected with a water quality sonde on Big Creek (RM 1.3; STORET: P09K06). Temperature (°C), D.O. (mg/L), pH (S.U.) and specific conductance (µS/cm) are included. The data was collected from 9/9-11/2015.

Figure 64: Plot of hourly data collected with a water quality sonde on Beaver Creek (RM 8.3; STORET: P10K03). Temperature (°C), D.O. (mg/L), pH (S.U.) and specific conductance (µS/cm) are included. The data was collected from 9/9-11/2015.
Figure 65: Plot of hourly data collected with a water quality sonde on Beaver Creek (RM 2.73; STORET: P10K02). Temperature (°C), D.O. (mg/L), pH (S.U.) and specific conductance (µS/cm) are included. The data was collected from 9/9-11/2015.

Figure 66: Plot of hourly data collected with a water quality sonde on Hammer Creek (RM 1.34; STORET: 303000). Temperature (°C), D.O. (mg/L), pH (S.U.) and specific conductance (µS/cm) are included. The data was collected from 9/9-11/2015.
Figure 67: Plot of hourly data collected with a water quality sonde on Jackson Cutoff Ditch (RM 1.15; STORET: 510040). Temperature (°C), D.O. (mg/L), pH (S.U.) and specific conductance (µS/cm) are included. The data was collected from 9/9-11/2015.

Figure 68: Plot of hourly data collected with a water quality sonde on Little Yellow Creek (RM 0.9; STORET: 500700). Temperature (°C), D.O. (mg/L), pH (S.U.) and specific conductance (µS/cm) are included. The data was collected from 9/9-11/2015.
Figure 69: Plot of hourly data collected with a water quality sonde on West Creek (trib to Yellow Ck) (RM 0.1; STORET: 302840). Temperature (°C), D.O. (mg/L), pH (S.U.) and specific conductance (µS/cm) are included. The data was collected from 9/9-11/2015.

Figure 70: Plot of hourly data collected with a water quality sonde on North Creek (RM 2.95; STORET: P06W17). Temperature (°C), D.O. (mg/L), pH (S.U.) and specific conductance (µS/cm) are included. The data was collected from 6/14-15/2016.
Figure 71: Plot of hourly data collected with a water quality sonde on North Creek (RM 2.6; STORET: P06K16). Temperature (°C), D.O. (mg/L), pH (S.U.) and specific conductance (µS/cm) are included. The data was collected from 6/14-15/2016.

Figure 72: Plot of hourly data collected with a water quality sonde on South Creek (RM 5.42; STORET: 302971). Temperature (°C), D.O. (mg/L), pH (S.U.) and specific conductance (µS/cm) are included. The data was collected from 6/14-15/2016.
Figure 73: Plot of hourly data collected with a water quality sonde on South Creek (RM 2.08; STORET: 302970). Temperature (°C), D.O. (mg/L), pH (S.U.) and specific conductance (µS/cm) are included. The data was collected from 6/14-15/2016.

Figure 74: Plot of hourly data collected with a water quality sonde on Platter Creek (RM 5.4; STORET: 302974). Temperature (°C), D.O. (mg/L), pH (S.U.) and specific conductance (µS/cm) are included. The data was collected from 6/14-15/2016.
Figure 75: Plot of hourly data collected with a water quality sonde on Snooks Run (RM 0.5; STORET: P06K17). Temperature (°C), D.O. (mg/L), pH (S.U.) and specific conductance (µS/cm) are included. The data was collected from 6/14-15/2016.

Figure 76: Plot of hourly data collected with a water quality sonde on Lost Creek (RM 1; STORET: P09S09). Temperature (°C), D.O. (mg/L), pH (S.U.) and specific conductance (µS/cm) are included. The data was collected from 6/14-15/2016.
Figure 77: Plot of hourly data collected with a water quality sonde on Brinkman Ditch (RM 2.8; STORET: P09K11). Temperature (°C), D.O. (mg/L), pH (S.U.) and specific conductance (µS/cm) are included. The data was collected from 6/14-15/2016.

Figure 78: Plot of hourly data collected with a water quality sonde on Brinkman Ditch (RM 2.35; STORET: P09W17). Temperature (°C), D.O. (mg/L), pH (S.U.) and specific conductance (µS/cm) are included. The data was collected from 6/14-15/2016.
Figure 79: Plot of hourly data collected with a water quality sonde on Little Yellow Creek (RM 4.6; STORET: P10W08). Temperature (°C), D.O. (mg/L), pH (S.U.) and specific conductance (µS/cm) are included. The data was collected from 6/14-15/2016.

Figure 80: Plot of hourly data collected with a water quality sonde on West Branch Tontogany River (RM 3.42; STORET: P10P13). Temperature (°C), D.O. (mg/L), pH (S.U.) and specific conductance (µS/cm) are included. The data was collected from 6/14-15/2016.
Figure 81: Plot of hourly data collected with a water quality sonde on West Branch Tontogany River (RM 2.19; STORET: P10P14). Temperature (°C), D.O. (mg/L), pH (S.U.) and specific conductance (µS/cm) are included. The data was collected from 6/14-15/2016.
Appendix F

QHEI Attributes Matrix
Table 1x. Matrix of habitat attributes and QHEI scores for sites sampled in the Maumee River Tributaries survey, 2015.

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<thead>
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<th>River Mile</th>
<th>Gradient (ft/mi)</th>
<th>Year</th>
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<td>5.56</td>
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</table>
Table 1. Matrix of habitat attributes and QHEI scores for sites sampled in the Maumee River Tributaries survey, 2015.

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<tr>
<th>River Mile</th>
<th>Gradient (ft/mi)</th>
<th>QHEI</th>
<th>WWH Attributes</th>
<th>MWH Attributes</th>
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Table 1x. Matrix of habitat attributes and QHEI scores for sites sampled in the Maumee River Tributaries survey, 2015.

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Key

QHEI Components

| Not Channelized or Recovered | Boulder/Cobble/Gravel Substrates | Split Fine Substrates | Good/Excellent Development | Moderate/High Sinuosity | Extensive/Moderate Cover | Fast Current/Eddies | Low/Normal Embayment | Max Depth >40cm | Low/Normal Riffle Embayment | Hi-influence Modified Attributes | MWH H.I.+1/WWH+1 Ratio | MWH M.I.+1/WWH+1 Ratio | EAS/2017-XX-XX Maumee River Tribuaries June 6, 2017 |
Appendix G

Surface Water Chemistry
Inorganic Parameters
Appendix Table 1. Inorganic chemistry and field parameter results for samples collected during the 2015 Maumee Tribs survey.

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### Appendix Table 1. Inorganic chemistry and field parameter results for samples collected during the 2015 Maumee Tribs survey.

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## Appendix Table 1. Inorganic chemistry and field parameter results for samples collected during the 2015 Maumee Tribs survey.

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Appendix Table 1. Inorganic chemistry and field parameter results for samples collected during the 2015 Maumee Tribs survey.

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### Appendix Table 1. Inorganic chemistry and field parameter results for samples collected during the 2015 Maumee Tribs survey.

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Appendix Table 1. Inorganic chemistry and field parameter results for samples collected during the 2015 Maumee Tribs survey.

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### Appendix Table 1. Inorganic chemistry and field parameter results for samples collected during the 2015 Maumee Tribs survey.

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<thead>
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<td>Barium ug/L</td>
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<td>Cadmium ug/L</td>
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<td>Calcium mg/L</td>
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<td>CBOD20 mg/L</td>
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<td>Chloride mg/L</td>
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<td>Manganese ug/L</td>
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<td>Spcond umhos/cm</td>
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<td>Zinc ug/L</td>
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Appendix Table 1. Inorganic chemistry and field parameter results for samples collected during the 2015 Maumee Tribs survey.

<table>
<thead>
<tr>
<th>Inorganic Parameters</th>
<th>6/15/15</th>
<th>7/13/15</th>
<th>7/27/15</th>
<th>8/10/15</th>
<th>8/24/15</th>
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<td>1001</td>
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<td>22.8</td>
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<td>0.42</td>
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<td>77</td>
<td>85</td>
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### Appendix Table 1. Inorganic chemistry and field parameter results for samples collected during the 2015 Maumee Tribs survey.

<table>
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<th>8/25/15</th>
<th>9/10/15</th>
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<tr>
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<td>10</td>
<td>&lt;5</td>
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Appendix Table 1. Inorganic chemistry and field parameter results for samples collected during the 2015 Maumee Tribs survey.

<table>
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<td>24</td>
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<td>&lt;5</td>
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Appendix Table 1. Inorganic chemistry and field parameter results for samples collected during the 2015 Maumee Tribs survey.

<table>
<thead>
<tr>
<th>Parameter</th>
<th>6/15/15</th>
<th>7/13/15</th>
<th>7/27/15</th>
<th>8/10/15</th>
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Appendix Table 1. Inorganic chemistry and field parameter results for samples collected during the 2015 Maumee Tribs survey.

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Appendix Table 1. Inorganic chemistry and field parameter results for samples collected during the 2015 Maumee Tribs survey.

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<th>6/15/15</th>
<th>7/13/15</th>
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<th>8/24/15</th>
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### Inorganic Parameters

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### Appendix Table 1. Inorganic chemistry and field parameter results for samples collected during the 2015 Maumee Tribs survey.

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Appendix Table 1. Inorganic chemistry and field parameter results for samples collected during the 2015 Maumee Tribs survey.

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Appendix Table 1. Inorganic chemistry and field parameter results for samples collected during the 2015 Maumee Tribs survey.

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### Appendix Table 1. Inorganic chemistry and field parameter results for samples collected during the 2015 Maumee Tribs survey.

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Appendix Table 1. Inorganic chemistry and field parameter results for samples collected during the 2015 Maumee Tribs survey.

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### Inorganic Parameters

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### Appendix Table 1. Inorganic chemistry and field parameter results for samples collected during the 2015 Maumee Tribs survey.

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## Appendix Table 1. Inorganic chemistry and field parameter results for samples collected during the 2015 Maumee Tribs survey.

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### Inorganic Parameters

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Appendix Table 1. Inorganic chemistry and field parameter results for samples collected during the 2015 Maumee Tribs survey.

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### Appendix Table 1. Inorganic chemistry and field parameter results for samples collected during the 2015 Maumee Tribs survey.

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Appendix Table 1. Inorganic chemistry and field parameter results for samples collected during the 2015 Maumee Tribs survey.

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Appendix Table 1. Inorganic chemistry and field parameter results for samples collected during the 2015 Maumee Tribs survey.

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**Appendix Table 1.** Inorganic chemistry and field parameter results for samples collected during the 2015 Maumee Tribs survey.

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Appendix Table 1. Inorganic chemistry and field parameter results for samples collected during the 2015 Maumee Tribs survey.

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Appendix Table 1. Inorganic chemistry and field parameter results for samples collected during the 2015 Maumee Tribs survey.

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Appendix Table 1. Inorganic chemistry and field parameter results for samples collected during the 2015 Maumee Tribs survey.

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### Appendix Table 1. Inorganic chemistry and field parameter results for samples collected during the 2015 Maumee Tribs survey.

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Appendix Table 1. Inorganic chemistry and field parameter results for samples collected during the 2015 Maumee Tribs survey.

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## Inorganic Parameters

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Appendix Table 1. Inorganic chemistry and field parameter results for samples collected during the 2015 Maumee Tribs survey.

<table>
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<tr>
<th>Inorganic Parameters</th>
<th>6/18/15</th>
<th>7/16/15</th>
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<th>8/13/15</th>
<th>8/27/15</th>
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<tbody>
<tr>
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## Inorganic Parameters

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### Appendix Table 1. Inorganic chemistry and field parameter results for samples collected during the 2015 Maumee Tribs survey.

**Station:** 303003  
**JACKSON CUTOFF DITCH DST YELLOW CREEK @ BAYS RD.**

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<td>.034</td>
<td>.051</td>
<td>.063</td>
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<td>66</td>
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<td>23</td>
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### Appendix Table 1. Inorganic chemistry and field parameter results for samples collected during the 2015 Maumee Tribs survey.

<table>
<thead>
<tr>
<th>Parameter</th>
<th>6/18/15</th>
<th>7/16/15</th>
<th>7/30/15</th>
<th>8/13/15</th>
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<tbody>
<tr>
<td>Alkalinity mg/L</td>
<td>268</td>
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<td>314</td>
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### Inorganic Parameters

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Appendix Table 1. Inorganic chemistry and field parameter results for samples collected during the 2015 Maumee Tribs survey.

| Station: 303006 TONTOGANY CREEK UPST TONTOGANY @ TONTOGANY RD. |
| River Code          | 04-013-000 | River Mile | 4.15 | Drainage Area | 11 | Lat   | 41.42366 | Long   | -83.73762 | Hydro Unit | 04100009-06-01 |

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## Appendix Table 1. Inorganic chemistry and field parameter results for samples collected during the 2015 Maumee Tribs survey.

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Appendix Table 1. Inorganic chemistry and field parameter results for samples collected during the 2015 Maumee Tribs survey.

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Appendix Table 1. Inorganic chemistry and field parameter results for samples collected during the 2015 Maumee Tribs survey.

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Appendix Table 1. Inorganic chemistry and field parameter results for samples collected during the 2015 Maumee Tribs survey.

### Inorganic Parameters

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Appendix Table 1. Inorganic chemistry and field parameter results for samples collected during the 2015 Maumee Tribs survey.

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### Appendix Table 1. Inorganic chemistry and field parameter results for samples collected during the 2015 Maumee Tribs survey.

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### Appendix Table 1. Inorganic chemistry and field parameter results for samples collected during the 2015 Maumee Tribs survey.

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## Inorganic Chemistry and Field Parameter Results

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Appendix Table 1. Inorganic chemistry and field parameter results for samples collected during the 2015 Maumee Tribs survey.

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## Appendix Table 1. Inorganic chemistry and field parameter results for samples collected during the 2015 Maumee Tribs survey.

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### Appendix Table 1. Inorganic chemistry and field parameter results for samples collected during the 2015 Maumee Tribs survey.

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Appendix Table 1. Inorganic chemistry and field parameter results for samples collected during the 2015 Maumee Tribs survey.

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<td>Calcium mg/L</td>
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<td>Chloride mg/L</td>
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<tr>
<td>Chlorophyll ug/L</td>
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<tr>
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<td>DO sat percent</td>
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<tr>
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<tr>
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<td>Phosphorus mg/L</td>
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<tr>
<td>Potassium mg/L</td>
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<td>Spcond umhos/cm</td>
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<td>Sulfate mg/L</td>
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<td>&lt;5</td>
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<tr>
<td>Zinc ug/L</td>
<td>&lt;10</td>
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</table>
Appendix Table 1. Inorganic chemistry and field parameter results for samples collected during the 2015 Maumee Tribs survey.

Station: P09K11  |  BRINKMAN DITCH UPST. HOLGATE WWTP @ CO. RD. 15
River Code  |  River Mile  | Drainage Area  | Lat  | Long  | Hydro Unit
04-036-000  |  2.80        |  8             | 41.26440 | -84.15220 | 04100009-01-03

<table>
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<tr>
<th>Inorganic Parameters</th>
<th>6/17/15</th>
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<tr>
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<td>.071</td>
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<td>37.6</td>
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<td>20</td>
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### Appendix Table 1. Inorganic chemistry and field parameter results for samples collected during the 2015 Maumee Tribs survey.

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<td>218</td>
<td>197</td>
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### Appendix Table 1. Inorganic chemistry and field parameter results for samples collected during the 2015 Maumee Tribs survey.

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Appendix Table 1. Inorganic chemistry and field parameter results for samples collected during the 2015 Maumee Tribs survey.

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### Appendix Table 1. Inorganic chemistry and field parameter results for samples collected during the 2015 Maumee Tribs survey.

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Appendix Table 1. Inorganic chemistry and field parameter results for samples collected during the 2015 Maumee Tribs survey.

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Appendix Table 1. Inorganic chemistry and field parameter results for samples collected during the 2015 Maumee Tribs survey.

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### Appendix Table 1. Inorganic chemistry and field parameter results for samples collected during the 2015 Maumee Tribs survey.

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Appendix Table 1. Inorganic chemistry and field parameter results for samples collected during the 2015 Maumee Tribs survey.

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### Appendix Table 1. Inorganic chemistry and field parameter results for samples collected during the 2015 Maumee Tribs survey.

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### Appendix Table 1. Inorganic chemistry and field parameter results for samples collected during the 2015 Maumee Tribs survey.

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Appendix Table 1. Inorganic chemistry and field parameter results for samples collected during the 2015 Maumee Tribs survey.

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### Appendix Table 1. Inorganic chemistry and field parameter results for samples collected during the 2015 Maumee Tribs survey.

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Appendix Table 1. Inorganic chemistry and field parameter results for samples collected during the 2015 Maumee Tribs survey.

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Appendix Table 1. Inorganic chemistry and field parameter results for samples collected during the 2015 Maumee Tribs survey.

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Appendix Table 1. Inorganic chemistry and field parameter results for samples collected during the 2015 Maumee Tribs survey.

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### Appendix Table 1. Inorganic chemistry and field parameter results for samples collected during the 2015 Maumee Tribs survey.

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<th>Parameter</th>
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## Appendix Table 1. Inorganic chemistry and field parameter results for samples collected during the 2015 Maumee Tribs survey.

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### Inorganic Parameters

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Appendix Table 1. Inorganic chemistry and field parameter results for samples collected during the 2015 Maumee Tribs survey.

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Appendix Table 1. Inorganic chemistry and field parameter results for samples collected during the 2015 Maumee Tribs survey.

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## Appendix Table 1. Inorganic chemistry and field parameter results for samples collected during the 2015 Maumee Tribs survey.

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**River Mile:** 2.73  
**Drainage Area:** 184  
**Lat:** 41.39360  
**Long:** -83.84500  
**Hydro Unit:** 04100009-05-09  

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Appendix Table 1. Inorganic chemistry and field parameter results for samples collected during the 2015 Maumee Tribs survey.

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### Appendix Table 1. Inorganic chemistry and field parameter results for samples collected during the 2015 Maumee Tribs survey.

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## Inorganic Parameters

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Appendix Table 1. Inorganic chemistry and field parameter results for samples collected during the 2015 Maumee Tribs survey.

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Appendix Table 1. Inorganic chemistry and field parameter results for samples collected during the 2015 Maumee Tribs survey.

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Appendix Table 1. Inorganic chemistry and field parameter results for samples collected during the 2015 Maumee Tribs survey.

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### Appendix Table 1. Inorganic chemistry and field parameter results for samples collected during the 2015 Maumee Tribs survey.

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<thead>
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<th>Parameter</th>
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Appendix Table 1. Inorganic chemistry and field parameter results for samples collected during the 2015 Maumee Tribs survey.

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Appendix Table 1. Inorganic chemistry and field parameter results for samples collected during the 2015 Maumee Tribs survey.

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## Appendix Table 1. Inorganic chemistry and field parameter results for samples collected during the 2015 Maumee Tribs survey.

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### Inorganic Parameters

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Note: Spcond umhos/cm and TSS mg/L values are not included in the table.
Appendix Table 1. Inorganic chemistry and field parameter results for samples collected during the 2015 Maumee Tribs survey.

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Appendix Table 1. Inorganic chemistry and field parameter results for samples collected during the 2015 Maumee Tribs survey.

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### Appendix Table 1. Inorganic chemistry and field parameter results for samples collected during the 2015 Maumee Tribs survey.

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### Appendix Table 1. Inorganic chemistry and field parameter results for samples collected during the 2015 Maumee Tribs survey.

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<td>22</td>
<td>23</td>
<td>24</td>
<td>21</td>
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<tr>
<td>Copper ug/L</td>
<td>6</td>
<td>3.3</td>
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<td>3.1</td>
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<tr>
<td>DO mg/L</td>
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<td>9.15</td>
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</tr>
<tr>
<td>DOsat percent</td>
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<td>93.4</td>
<td>98.2</td>
<td>87</td>
<td>99</td>
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</tr>
<tr>
<td>Hardness mg/L</td>
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<td>315</td>
<td>339</td>
<td>318</td>
<td>325</td>
<td></td>
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<tr>
<td>Iron ug/L</td>
<td>3400</td>
<td>771</td>
<td>377</td>
<td>885</td>
<td>263</td>
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<tr>
<td>Lead ug/L</td>
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<td>&lt;2</td>
<td>&lt;2</td>
<td>.5</td>
<td>.2</td>
<td></td>
</tr>
<tr>
<td>Magnesium mg/L</td>
<td>18.4</td>
<td>19.5</td>
<td>22.6</td>
<td>21</td>
<td>23.1</td>
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<tr>
<td>Manganese ug/L</td>
<td>83</td>
<td>40</td>
<td>23</td>
<td>40</td>
<td>21</td>
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<tr>
<td>Nickel ug/L</td>
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<td>4.3</td>
<td>4.1</td>
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<tr>
<td>Nitrate+Nitrite mg/L</td>
<td>19.6</td>
<td>13.4</td>
<td>9.9</td>
<td>7.15</td>
<td>5.89</td>
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<tr>
<td>Nitrite mg/L</td>
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<td>.045</td>
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<td>.018</td>
<td>.014</td>
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<td>7.67</td>
<td>8.1</td>
<td>8.14</td>
<td>8.19</td>
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<tr>
<td>Pheophytin ug/L</td>
<td></td>
<td></td>
<td></td>
<td>1.4</td>
<td></td>
<td></td>
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<tr>
<td>Phosphate mg/L</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Phosphorus mg/L</td>
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<td>.042</td>
<td>.04</td>
<td>.035</td>
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<td>Potassium mg/L</td>
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<td>8.4</td>
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<td>9.9</td>
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<tr>
<td>Selenium ug/L</td>
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<td>&lt;2</td>
<td>&lt;2</td>
<td>.7</td>
<td>1</td>
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<tr>
<td>Sodium mg/L</td>
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<td>18.1</td>
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<td>Spcond umhos/cm</td>
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<td>695</td>
<td>748</td>
<td>805</td>
<td>778</td>
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</tr>
<tr>
<td>Strontium mg/L</td>
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<td>412</td>
<td>464</td>
<td>478</td>
<td>480</td>
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<tr>
<td>Sulfate mg/L</td>
<td>35.2</td>
<td>35.1</td>
<td>43.4</td>
<td>41</td>
<td>49.4</td>
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<tr>
<td>TDS mg/L</td>
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<td>442</td>
<td>478</td>
<td>460</td>
<td>486</td>
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</tr>
<tr>
<td>TempC deg C</td>
<td>19.3</td>
<td>19.2</td>
<td>21.9</td>
<td>21.4</td>
<td>19.1</td>
<td></td>
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<tr>
<td>TKN mg/L</td>
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<tr>
<td>TSS mg/L</td>
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<td>18</td>
<td>10</td>
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<tr>
<td>Zinc ug/L</td>
<td>19</td>
<td>&lt;10</td>
<td>&lt;10</td>
<td>4</td>
<td>4</td>
<td></td>
</tr>
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</table>

Detected in Blank; < Exceeds Calibration; > QC Criteria Not Met; Invalid Colony Count; CoAnalyeCorrelation; Matrix Interference; Estimated Value; Holding/Shipping Time Exceeded; PesticideGCDiff
## Appendix Table 1. Inorganic chemistry and field parameter results for samples collected during the 2015 Maumee Tribs survey.

<table>
<thead>
<tr>
<th>Parameter</th>
<th>9/9/15</th>
<th>9/10/15</th>
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<tbody>
<tr>
<td><strong>Alkalinity mg/L</strong></td>
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<tr>
<td><strong>Aluminum ug/L</strong></td>
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<td>233</td>
</tr>
<tr>
<td><strong>Ammonia mg/L</strong></td>
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<td>.079</td>
</tr>
<tr>
<td><strong>Arsenic ug/L</strong></td>
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<td>2.3</td>
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<tr>
<td><strong>Barium ug/L</strong></td>
<td>86</td>
<td>82</td>
</tr>
<tr>
<td><strong>Cadmium ug/L</strong></td>
<td>&lt;.2</td>
<td>&lt;.2</td>
</tr>
<tr>
<td><strong>Calcium mg/L</strong></td>
<td>83.8</td>
<td>83</td>
</tr>
<tr>
<td><strong>CBOD20 mg/L</strong></td>
<td>6.9</td>
<td></td>
</tr>
<tr>
<td><strong>Chloride mg/L</strong></td>
<td>84.4</td>
<td>80.2</td>
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<tr>
<td><strong>Chlorophyll ug/L</strong></td>
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<td></td>
</tr>
<tr>
<td><strong>Chromium ug/L</strong></td>
<td>&lt;2</td>
<td>&lt;2</td>
</tr>
<tr>
<td><strong>COD mg/L</strong></td>
<td></td>
<td>26</td>
</tr>
<tr>
<td><strong>Copper ug/L</strong></td>
<td>2.5</td>
<td>2.7</td>
</tr>
<tr>
<td><strong>DO mg/L</strong></td>
<td>7.97</td>
<td>7.22</td>
</tr>
<tr>
<td><strong>DOsat percent</strong></td>
<td>96</td>
<td>82.2</td>
</tr>
<tr>
<td><strong>Hardness mg/L</strong></td>
<td>319</td>
<td>310</td>
</tr>
<tr>
<td><strong>Iron ug/L</strong></td>
<td>340</td>
<td>414</td>
</tr>
<tr>
<td><strong>Lead ug/L</strong></td>
<td>&lt;2</td>
<td>&lt;2</td>
</tr>
<tr>
<td><strong>Magnesium mg/L</strong></td>
<td>26.7</td>
<td>25</td>
</tr>
<tr>
<td><strong>Manganese ug/L</strong></td>
<td>50</td>
<td>59</td>
</tr>
<tr>
<td><strong>Nickel ug/L</strong></td>
<td>3.8</td>
<td>3.9</td>
</tr>
<tr>
<td><strong>Nitrate+Nitrite mg/L</strong></td>
<td>2.52</td>
<td>2.21</td>
</tr>
<tr>
<td><strong>Nitrite mg/L</strong></td>
<td>.065</td>
<td>.06</td>
</tr>
<tr>
<td><strong>pH su</strong></td>
<td>8.12</td>
<td>8.09</td>
</tr>
<tr>
<td><strong>Pheophytin ug/L</strong></td>
<td>2.3</td>
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</tr>
<tr>
<td><strong>Phosphate mg/L</strong></td>
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<tr>
<td><strong>Phosphorus mg/L</strong></td>
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<td>.041</td>
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<tr>
<td><strong>Potassium mg/L</strong></td>
<td>11.5</td>
<td>11.4</td>
</tr>
<tr>
<td><strong>Selenium ug/L</strong></td>
<td>&lt;2</td>
<td>&lt;2</td>
</tr>
<tr>
<td><strong>Sodium mg/L</strong></td>
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<td>41.4</td>
</tr>
<tr>
<td><strong>Spcond umhos/cm</strong></td>
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<td>798</td>
</tr>
<tr>
<td><strong>Strontium ug/L</strong></td>
<td>520</td>
<td>505</td>
</tr>
<tr>
<td><strong>Sulfate mg/L</strong></td>
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<td>57</td>
</tr>
<tr>
<td><strong>TDS mg/L</strong></td>
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<td>506</td>
</tr>
<tr>
<td><strong>TempC deg C</strong></td>
<td>24.6</td>
<td>21.7</td>
</tr>
<tr>
<td><strong>TKN mg/L</strong></td>
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<td>1.22</td>
</tr>
<tr>
<td><strong>TSS mg/L</strong></td>
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<td>11</td>
</tr>
<tr>
<td><strong>Zinc ug/L</strong></td>
<td>&lt;10</td>
<td>&lt;10</td>
</tr>
</tbody>
</table>
Organic Parameters
## Appendix Table 2. Results from organic scans of water column samples collected during the 2015 Maumee Tribs survey.

All units are ug/L.

<table>
<thead>
<tr>
<th>Rivercode</th>
<th>River Mile</th>
<th>Drainage Area</th>
<th>Hydro Unit</th>
<th>Lat</th>
<th>Long</th>
</tr>
</thead>
<tbody>
<tr>
<td>04-037-001</td>
<td>1.02</td>
<td>9.9</td>
<td>04100009-04-02</td>
<td>41.44578</td>
<td>-84.05721</td>
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</tbody>
</table>

<table>
<thead>
<tr>
<th>Substance</th>
<th>Value</th>
</tr>
</thead>
<tbody>
<tr>
<td>4-Chloro-3-methylphenol</td>
<td>&lt;10.8</td>
</tr>
<tr>
<td>Acenaphthene</td>
<td>&lt;5.4</td>
</tr>
<tr>
<td>Acenaphthylene</td>
<td>&lt;5.4</td>
</tr>
<tr>
<td>Anthracene</td>
<td>&lt;2.2</td>
</tr>
<tr>
<td>Benzo[a]anthracene</td>
<td>&lt;2.2</td>
</tr>
<tr>
<td>Benzo[a]pyrene</td>
<td>&lt;2.2</td>
</tr>
<tr>
<td>Benzo[b]fluoranthene</td>
<td>&lt;2.2</td>
</tr>
<tr>
<td>Benzo[g,h,i]perylene</td>
<td>&lt;2.2</td>
</tr>
<tr>
<td>Benzo[k]fluoranthene</td>
<td>&lt;2.2</td>
</tr>
<tr>
<td>bis(2-chloroethoxy) methane</td>
<td>&lt;5.4</td>
</tr>
<tr>
<td>bis(2-chloroethyl) ether</td>
<td>&lt;2.2</td>
</tr>
<tr>
<td>bis(2-Chloroisopropyl) ether</td>
<td>&lt;2.2</td>
</tr>
<tr>
<td>bis(2-ethylhexyl) phthalate (DEHP)</td>
<td>&lt;10.8</td>
</tr>
<tr>
<td>bis(n-octyl) phthalate</td>
<td>&lt;2.2</td>
</tr>
<tr>
<td>Bromophenyl-4 phenyl ether</td>
<td>&lt;5.4</td>
</tr>
<tr>
<td>Butyl benzyl phthalate</td>
<td>&lt;2.2</td>
</tr>
<tr>
<td>Chloronaphthalene-2</td>
<td>&lt;5.4</td>
</tr>
<tr>
<td>Chlorophenol-2</td>
<td>&lt;2.2</td>
</tr>
<tr>
<td>Chlorophenyl-4 phenyl ether</td>
<td>&lt;2.2</td>
</tr>
<tr>
<td>Chrysene</td>
<td>&lt;2.2</td>
</tr>
<tr>
<td>Dibenzo[a,h]anthracene</td>
<td>&lt;2.2</td>
</tr>
<tr>
<td>Dibutyl phthalate</td>
<td>&lt;5.4</td>
</tr>
<tr>
<td>Dichlorobenzene, 1,2-</td>
<td>&lt;2.2</td>
</tr>
<tr>
<td>Dichlorobenzene, 1,3-</td>
<td>&lt;2.2</td>
</tr>
<tr>
<td>Dichlorobenzene, 1,4-</td>
<td>&lt;2.2</td>
</tr>
<tr>
<td>Dichlorophenol, 2,4-</td>
<td>&lt;2.2</td>
</tr>
<tr>
<td>Diethyl phthalate</td>
<td>&lt;5.4</td>
</tr>
<tr>
<td>Dimethyl phthalate</td>
<td>&lt;5.4</td>
</tr>
<tr>
<td>Dimethylphenol, 2,4-</td>
<td>&lt;10.8</td>
</tr>
<tr>
<td>Dinitro-o-cresol</td>
<td>&lt;5.4</td>
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<tr>
<td>Dinitrophenol, 2,4-</td>
<td>&lt;21.5</td>
</tr>
<tr>
<td>Dinitrotoluene, 2,4-</td>
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<tr>
<td>Dinitrotoluene, 2,6-</td>
<td>&lt;2.2</td>
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<tr>
<td>Fluoranthene</td>
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<td>Fluorene</td>
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<tr>
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<td>Hexachlorobutadiene</td>
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<td>Hexachlorocyclopentadiene</td>
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<td>Isophorone</td>
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<td>Nitrophenol, 2-</td>
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<td>Nitrophenol, 4-</td>
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<td>Nitrosodiphenylamine, n-</td>
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</table>
Appendix Table 2. Results from organic scans of water column samples collected during the 2015 Maumee Tribs survey. All units are ug/L.

<table>
<thead>
<tr>
<th>Rivercode</th>
<th>River Mile</th>
<th>Drainage Area</th>
<th>Hydro Unit</th>
<th>Lat</th>
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<td>9.9</td>
<td>04100009-04-02</td>
<td>41.44578</td>
<td>-84.05721</td>
</tr>
</tbody>
</table>

| 2015/6/16 | Nitrosodipropylamine, n- | <2.2 |
| Pentachlorophenol (PCP) | <10.8 |
| Phenanthrene | <2.2 |
| Phenol | <2.2 |
| Pyrene | <2.2 |
| Trichlorobenzene, 1,2,4- | <2.2 |
| Trichlorophenol, 2,4,6- (TCPh) | <5.4 |
### Appendix Table 2. Results from organic scans of water column samples collected during the 2015 Maumee Tribs survey.

All units are ug/L.

<table>
<thead>
<tr>
<th>Rivercode</th>
<th>River Mile</th>
<th>Drainage Area</th>
<th>Hydro Unit</th>
<th>Lat</th>
<th>Long</th>
</tr>
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*2015/6/18*

<table>
<thead>
<tr>
<th>Compound</th>
<th>Concentration</th>
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<tbody>
<tr>
<td>4-Chloro-3-methylphenol</td>
<td>&lt;10.9</td>
</tr>
<tr>
<td>Acenaphthene</td>
<td>&lt;5.5</td>
</tr>
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<td>&lt;5.5</td>
</tr>
<tr>
<td>Chlorophenol-2</td>
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<tr>
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<td>&lt;2.2</td>
</tr>
<tr>
<td>Chrysene</td>
<td>&lt;2.2</td>
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<tr>
<td>Dibenzo[a,h]anthracene</td>
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<tr>
<td>Dibutyl phthalate</td>
<td>&lt;5.5</td>
</tr>
<tr>
<td>Dichlorobenzene, 1,2-</td>
<td>&lt;2.2</td>
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<tr>
<td>Dichlorobenzene, 1,3-</td>
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<td>Dichlorobenzene, 1,4-</td>
<td>&lt;2.2</td>
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<tr>
<td>Dichlorophenol, 2,4-</td>
<td>&lt;2.2</td>
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<tr>
<td>Diethyl phthalate</td>
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<tr>
<td>Dimethylphenol, 2,4-</td>
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<tr>
<td>Dinitro-o cresol</td>
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<tr>
<td>Dinitrophenol, 2,4-</td>
<td>&lt;21.9</td>
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<tr>
<td>Dinitrotoluene, 2,4-</td>
<td>&lt;2.2</td>
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<tr>
<td>Dinitrotoluene, 2,6-</td>
<td>&lt;2.2</td>
</tr>
<tr>
<td>Fluoranthene</td>
<td>&lt;2.2</td>
</tr>
<tr>
<td>Fluorene</td>
<td>&lt;2.2</td>
</tr>
<tr>
<td>Hexachlorobenzene</td>
<td>&lt;2.2</td>
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<tr>
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<th>Long</th>
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All units are ug/L.

| Nitrosodipropylamine, n-     | <2.2 |
| Pentachlorophenol (PCP)      | <10.9 |
| Phenanthrene                 | <2.2 |
| Phenol                       | <2.2 |
| Pyrene                       | <2.2 |
| Trichlorobenzene, 1,2,4-     | <2.2 |
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2015/6/16
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<td>3.47</td>
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<td>-83.88192</td>
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2015/6/18

Nitrosodipropylamine, n-<2.2
Pentachlorophenol (PCP)<10.9
Phenanthrene<2.2
Phenol<2.2
Pyrene<2.2
Trichlorobenzene, 1,2,4-<2.2
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<td>-83.98097</td>
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Appendix Table 2. Results from organic scans of water column samples collected during the 2015 Maumee Tribs survey.

All units are ug/L.

<table>
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<tr>
<th>Rivercode</th>
<th>River Mile</th>
<th>Drainage Area</th>
<th>Hydro Unit</th>
<th>Lat</th>
<th>Long</th>
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<td>Pyrene</td>
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2015/6/16
Appendix Table 2. Results from organic scans of water column samples collected during the 2015 Maumee Tribs survey. All units are ug/L.

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* Detected in Blank; < Exceeds Calibration; ≥ QC Criteria Not Met; ‡ Invalid Colony Count; § CoAnalyteCorrelation; ‖ Matrix Interference; ¶ Estimated Value; ℄ Holding/Shipping Time Exceeded; # PesticideGCDiff
Appendix H

Sediment Chemistry
## Appendix Table 4. Organic parameters from sediment samples collected during the 2015 Maumee Tributaries survey.

All units are mg/kg.

<table>
<thead>
<tr>
<th>PARAMETER</th>
<th>RESULT</th>
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<th>PARAMETER</th>
<th>RESULT</th>
<th>normalized</th>
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<td>Pentachlorobenzene</td>
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All units are mg/kg.
Appendix Table 4. Organic parameters from sediment samples collected during the 2015 Maumee Tributaries survey.

All units are mg/kg.

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<th>PARAMETER</th>
<th>RESULT</th>
<th>normalized PAH</th>
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<td>bis(2-Chloroethoxy)methane</td>
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Appendix Table 4. Organic parameters from sediment samples collected during the 2015 Maumee Tributaries survey.

All units are mg/kg.

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Appendix Table 4. Organic parameters from sediment samples collected during the 2015 Maumee Tributaries survey.

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All units are mg/kg.
Appendix Table 4. Organic parameters from sediment samples collected during the 2015 Maumee Tributaries survey.

All units are mg/kg.  

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Detected in Blank; * Exceeds Calibration; † QC Criteria Not Met; ‡ Invalid Colony Count; †† CoAnalyteCorrelation; †‡ Matrix Interference; †§ Estimated Value; †‖ Holding/Shipping Time Exceeded; †¶ PesticideGCdiff
Appendix I

Recreation Use Sample Summary
| Station I.D. | Sample Location | Recreational Designation | E. coli | RL
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<td>302977</td>
<td>PRESTON RUN @ STANLEY RD.</td>
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<td>302981</td>
<td>PLATTER CREEK @ BERCHE RD. (ON CROSSING)</td>
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<td>P06417</td>
<td>DOWNS RUN AT THE Bend @ SLOUGH RD.</td>
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<td>P0624</td>
<td>RABIE DELAMERE CREEK NEAR MOUTH @ TWP. RD. 150</td>
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<td>GORDON CREEK SW OF SHERWOOD @ COUNTY LINE RD. (LOWER CROSSING)</td>
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<td>MILL CREEK NEAR MOUTH, 901 FOUNTAIN STREET RD.</td>
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<td>NORTH CREEK DST. ANTERWEP WWTP @ MURPHY RD.</td>
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<td>SULPHUR CREEK DST. SHERWOOD @ ROLAND RD</td>
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<td>BERNIE CREEK @ CR 7-C</td>
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<td>UT to Maumee River at MM 48.7 @ T-16</td>
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<td>VN MINING CREEK IN NAPOLEON @ OAKWOOD AVE</td>
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<td>302988</td>
<td>N. TURKEYFOOT CREEK @ CR 5</td>
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<td>OBERHAUS CREEK NEAR LIBERTY CENTER @ C-5</td>
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<td>KONZEN DITCH @ CR 10 site moved to P09K14</td>
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<td>KONZEN DITCH NEAR MOUTH @ CR 5</td>
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<td>S. TURKEYFOOT NW OF HAMLER @ CR 10</td>
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<td>UT TO BEAVER CREEK UPS HERTFILD FARM @ PO-20</td>
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<td>UT TO W BR TONTOGANY CREEK SS OF LONG-JUDSON</td>
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<td>YELLOW CREEK SS OF DEISLER @ CR 10</td>
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<td>BEAVER CREEK SS OF DESHLER @ HENRY/WOOD CO. LINE</td>
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<td>W. BR. TONTOGANY CREEK SS OF TONTOGANY @ TULLER</td>
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**Notes:**
- E. coli result was reported as "Not detected".
- Method reporting level (RL) is 2 colonies per 100 ml.

**Geometric Mean (GUM) for recreational criteria:**
- B: Background
- x: Undesignated